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(72) Inventors: CONTI, Marco; 23 Ryan Court, Stanford, CA  
94305 (US). JAISWAL, Bijay, Shankar; 189 Buckthorn  
Way, Menlo Park, CA 94025 (US).

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(74) Agent: BORDEN, Paula, A.; Bozicevic, Field & Francis  
LLP, Suite 200, 200 Middlefield Road, Menlo Park, CA  
94025 (US).

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(71) Applicant: THE BOARD OF TRUSTEES OF THE LE-  
LAND STANFORD JUNIOR UNIVERSITY [US/US];  
900 Welch Road, Suite 350, Palo Alto, CA 94304 (US).



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(54) Title: POLYNUCLEOTIDES ENCODING HUMAN SOLUBLE ADENYLYL CYCLASE, POLYPEPTIDES ENCODED  
THEREBY, AND METHODS OF USE THEREOF

(57) Abstract: The invention relates to compositions for and methods of reducing the number of motile sperm in a male. The invention provides methods for identifying substances which inhibit soluble adenylyl cyclase, and which therefore have potential as male contraceptives. The invention further provides isolated polynucleotide sequences encoding human soluble adenylyl cyclase, as well as vectors and host cells comprising the polynucleotide sequences. Further provided are isolated human sAC polypeptides.

**POLYNUCLEOTIDES ENCODING HUMAN SOLUBLE ADENYLYL CYCLASE,  
POLYPEPTIDES ENCODED THEREBY, AND METHODS OF USE THEREOF**

**GOVERNMENT RIGHTS**

5 The United States Government may have certain rights in this application pursuant to  
National Institutes of Health Grant No. HD31544.

**FIELD OF THE INVENTION**

This application is in the field of male contraception. More particularly, the invention relates  
10 to inhibition of a soluble adenylyl cyclase.

**BACKGROUND OF THE INVENTION**

Presently, the major burden of fertility regulation is carried by women, since most available  
methods are female methods. Oral contraceptives, implants and injectables, intrauterine devices, tubal  
15 ligation, diaphragms, and spermicides are all methods practiced by women. Methods of male  
contraception currently available or being developed include vasectomy, condoms, and hormonal  
methods.

Spermatogenesis is a differentiation process whereby male germ cells develop into mature  
spermatozoa. Leblond, et al. (1952) *Ann. N.Y. Acad. Sci.* 55:548-573; Parvinen (1982) *Endocr. Rev.*  
20 3:404-417. Primordial germ cells, derived from primitive ectoderm, are established in the primitive  
gonad on embryonic day 10.5 in the mouse. After birth, these cells proliferate extensively, giving rise  
to type A spermatogonia which can either replicate as stem cells or differentiate to type B  
spermatogonia. At puberty, type B spermatogonia develop into large diploid primary spermatocytes  
that undergo two reductive divisions, giving rise to the haploid spermatids. Spermatids evolve into  
25 motile spermatozoa through a process referred to as spermiogenesis, characterized by restructuring of  
their nuclei and development of flagella.

The cyclic nucleotide-dependent pathway is thought to play an important role in the final  
maturation of spermatids. Mice deficient in cAMP responsive element modulator (CREM), a  
transacting factor downstream from the cAMP-dependent pathway, are infertile. CREM-null mice  
30 display a spermatogenic arrest at the initial spermatid stage, and no spermatozoa are produced. Nantel  
et al. (1996) *Nature* 380:159-162; and Blendy et al. (1996) *Nature* 380:162-165. Rather than  
differentiating into spermatozoa, spermatids lacking CREM undergo programmed cell death.  
Oligoazoospermic men were also found to have a disruption in CREM expression. Lin et al. (1998)  
35 *Fertil. Steril.* 69:533-538; and Weinbauer et al. (1998) *Mol. Hum. Reprod.* 4:9-15. Among patients  
with predominant round spermatid maturation arrest, CREM expression is significantly reduced or

undetectable, as revealed by quantitative polymerase chain reaction (PCR) analysis. CREM-negative spermatids fail to progress beyond stage III of spermatogenesis.

Male contraceptive methods that interrupt sperm transport in the male reproductive tract, are not without their complications or long term risks. Comhaire (1994) *Hum. Reprod.* 9:586-590. More complex approaches, such as regimens for the hormonal control of male fertility, have also not been fully satisfactory. Such methods have focused on the suppression of spermatogenesis to the point of azoospermia, a goal which has been difficult to achieve. Baird and Glasier (1993) *N. Engl. J. Med.* 328:1543-1549. This approach, nonetheless, is at the forefront of male contraceptive research, and awaits developments in the pharmacology of oral gonadotropin releasing hormone (GnRH) antagonists before its acceptability can be further advanced. Tom et al. (1992) *J. Clin. Endocrinol. Metab.* 10:476-483.

Other pharmacologic approaches to male contraception have studied the effects of various chemical agents on the functioning of the male reproductive tract. Unfortunately, these studies have not advanced much beyond the search for animal models, since the various side effects of chemicals tested make clinical testing with human males unrealistic. Zaneveld and Waller (1989) *Prog. Clin. Biol. Res.* 302:129-156. The use of the immune response to block contraception has been an important front in efforts to develop more sophisticated contraceptive systems. Unfortunately, such approaches have thus far failed, for a number of reasons. First, male autoimmunity against sperm does not suppress sperm production in men; this is known because such autoimmunity can occur after vasectomy. In addition, it has been found that female immunity against sperm does not necessarily result in infertility. Second, attempts to define the antigenic character of the human sperm surface are still in their infancy, and though epitopes have been identified on human sperm which may have contraceptive potential (Zhang et al. (1992) *Chin. Med. J.* 105:998-1033; Naz (1987) *J. Clin. Invest.* 80:1375-1383), the chemical identity and functions of these antigens themselves are unknown.

It is evident from the foregoing discussion that there is a need for methods of male contraception that are reversible, reliable, and avoid the aforementioned drawbacks. The present invention addresses this need and provides related advantages as well.

#### SUMMARY OF THE INVENTION

The present invention relates to the use of testis-specific form of adenylyl cyclase, referred to herein as sAC, as a target for male contraception. Inhibition of sAC enzyme activity provides a means for reversibly reducing the number of motile sperm in males, e.g., by inhibiting spermatogenesis, or by reversibly reducing or inhibiting sperm motility, and thus for effecting male contraception. Accordingly, the present invention provides methods for reducing the number of motile sperm in a male. In some embodiments, methods are provided for reducing the number of motile sperm in a

human male. In other embodiments, methods are provided for reducing the number of motile sperm in a non-human male, e.g., in rodent populations. In some embodiments, the present invention provides methods for reducing or inhibiting spermatogenesis in a male. In other embodiments, methods are provided for reducing or inhibiting sperm motility in a male. The invention further provides methods 5 for reversibly achieving male contraception. These methods generally comprising administering to a male a composition comprising an effective amount of a substance which inhibits or reduces sAC enzyme activity.

The invention further provides isolated human sAC polypeptides. Isolated sAC polypeptides are useful for detecting compounds that modulate sAC enzyme activity. Accordingly, the invention 10 further provides methods of identifying substances which modulate sAC enzyme activity. In these methods, sAC polypeptides of any species can be used. In some embodiments, the methods use a human sAC polypeptide and identify compounds that modulate sAC enzyme activity, particularly human sAC enzyme activity. In other embodiments, the methods use rat sAC polypeptides and identify compounds that modulate sAC enzyme activity, particularly rat sAC enzyme activity. The 15 methods generally comprise contacting an sAC enzyme with a substance to be tested, and determining an effect, if any, on sAC enzyme activity. The invention further provides compounds which modulate sAC activity.

The present invention further provides isolated polynucleotides encoding a human soluble adenylyl cyclase (human sAC), and further provides isolated human adenylyl cyclase polypeptides. 20 Polynucleotides and polypeptides of the invention find use in a variety of applications. Isolated human sAC polynucleotides are useful for producing human sAC polypeptides, for identifying modifications in sAC polynucleotides, and for detecting the presence of related polynucleotides. The invention further provides recombinant vectors and isolated host cells comprising polynucleotides of the invention.

25 These and other objects, advantages, and features of the invention will become apparent to those persons skilled in the art upon reading the details of the polynucleotides, polypeptides, and methods of the invention, as more fully described below.

#### BRIEF DESCRIPTION OF THE DRAWINGS

30 Figure 1 provides the amino acid sequence of human soluble adenylyl cyclase (sAC) (SEQ ID NO:2).

Figures 2A and 2B show an alignment of the amino acid sequences of human and rat sAC (SEQ ID NO 4).

35 Figures 3A and 3B provide a nucleotide sequence encoding human soluble adenylyl cyclase (SEQ ID NO:1).

Figures 4A-E show an alignment of the nucleotide sequence of exons encoding human sAC with a rat sAC nucleotide sequence (GenBank Accession No. AF081941; SEQ ID NO:3).

Figure 5 is a bar graph depicting activation of sAC adenylyl cyclase activity by bicarbonate.

5

## MODES OF CARRYING OUT THE INVENTION

The invention provides methods of detecting substances that modulate soluble adenylyl cyclase (sAC) enzyme activity. In these methods, the sAC can be from any species, including mammalian species. Compositions comprising effective amounts of the substances identified by these methods have the potential to reduce the number of motile sperm, e.g., by inhibiting spermatogenesis or reducing sperm motility, and therefore are of interest as male contraceptives. The invention further provides methods of reducing the number of motile sperm in a male. The invention further provides methods of inhibiting spermatogenesis, as well as methods of reducing sperm motility, in a male. The compositions and methods of the present invention provide means for human male contraception, as well as means for controlling animal populations, e.g., undesired rodent populations.

15 sAC is a cytosolic enzyme found in testis, particularly in spermatids, but not in somatic cells. This enzyme catalyzes the production of adenosine 3',5' cyclic monophosphate (cAMP). cAMP is an important mediator of spermatid differentiation. Accordingly, sAC is a target for substances which inhibit or reduce spermatogenesis, or reduce sperm motility, and which therefore have the potential to function as male contraceptives.

20 sAC, as described herein, is characterized by one or more of the following features: (1) it is expressed in high levels in the testis, particularly in spermatids, and, is either absent from, or is present at only low levels in somatic cells; (2) unlike membrane-associated adenylyl cyclases, it is not associated with G proteins, and thus is insensitive to G protein regulation (e.g., it is not activatable by forskolin); (3) it is found in the cytosol (i.e., is not an integral membrane protein); (4) its reaction 25 product is cAMP; (5) it exhibits an approximately 10-fold lower affinity for its substrate, adenosine triphosphate (ATP) than membrane-associated adenylyl cyclases ( $K_m \approx 1 \text{ mM}$  for sAC, compared with  $K_m \approx 100 \mu\text{M}$  for membrane-associated adenylyl cyclases); (6) its adenylyl cyclase activity is dependent on the presence of divalent manganese or magnesium ions; and (7) its adenylyl cyclase activity is activated by bicarbonate ions, e.g., its adenylyl cyclase activity is activated by sodium 30 bicarbonate.

The present invention further provides isolated polynucleotides comprising sequences which encode human soluble adenylyl cyclase (human sAC). Isolation of a human polynucleotide encoding sAC, and determination of its sequence, allows production of human sAC polypeptides.

Before the present invention is described, it is to be understood that this invention is not 35 limited to particular embodiments described, as such may, of course, vary. It is also to be understood

that the terminology used herein is for the purpose of describing particular embodiments only, and is not intended to be limiting, since the scope of the present invention will be limited only by the appended claims.

Where a range of values is provided, it is understood that each intervening value, to the tenth of the unit of the lower limit unless the context clearly dictates otherwise, between the upper and lower limits of that range is also specifically disclosed. Each smaller range between any stated value or intervening value in a stated range and any other stated or intervening value in that stated range is encompassed within the invention. The upper and lower limits of these smaller ranges may independently be included or excluded in the range, and each range where either, neither or both limits are included in the smaller ranges is also encompassed within the invention, subject to any specifically excluded limit in the stated range. Where the stated range includes one or both of the limits, ranges excluding either or both of those included limits are also included in the invention.

Unless defined otherwise, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs. Although any methods and materials similar or equivalent to those described herein can be used in the practice or testing of the present invention, the preferred methods and materials are now described. All publications mentioned herein are incorporated herein by reference to disclose and describe the methods and/or materials in connection with which the publications are cited.

It must be noted that as used herein and in the appended claims, the singular forms "a", "and", and "the" include plural referents unless the context clearly dictates otherwise. Thus, for example, reference to "an agent" includes a plurality of such agents and reference to "the polynucleotide" includes reference to one or more polynucleotides and equivalents thereof known to those skilled in the art, and so forth.

The publications discussed herein are provided solely for their disclosure prior to the filing date of the present application. Nothing herein is to be construed as an admission that the present invention is not entitled to antedate such publication by virtue of prior invention. Further, the dates of publication provided may be different from the actual publication dates which may need to be independently confirmed.

30 General Methods

The practice of the present invention will employ, unless otherwise indicated, conventional techniques of molecular biology, microbiology, recombinant DNA, and immunology, which are within the skill of the art. Such techniques are explained fully in the literature. *See e.g.*, Sambrook, Fritsch, and Maniatis, Molecular Cloning: A Laboratory Manual, Second Edition (1989), Oligonucleotide Synthesis (M. J. Gait Ed., 1984), Animal Cell Culture (R. I. Freshney, Ed., 1987), the series Methods

in Enzymology Academic Press, Inc.); Gene Transfer Vectors for Mammalian Cells (J. M. Miller and M. P. Calos Eds. 1987), Handbook of Experimental Immunology, (D. M. Weir and C. C. Blackwell, Eds.); Current Protocols in Molecular Biology, (F. M. Ausubel, et al., Eds. 1987, and updates); and Current Protocols in Immunology (J. E. Coligan, A. M. Kruisbeek, D. H. Margulies, E. M. Shevach and W. Strober Eds. 1991).

### Definitions

The terms "polynucleotide" and "nucleic acid", used interchangeably herein, refer to a polymeric forms of nucleotides of any length, either ribonucleotides or deoxynucleotides. Thus, this term includes, but is not limited to, single-, double-, or multi-stranded DNA or RNA, genomic DNA, cDNA, DNA-RNA hybrids, or a polymer comprising purine and pyrimidine bases or other natural, chemically or biochemically modified, non-natural, or derivatized nucleotide bases. The backbone of the polynucleotide can comprise sugars and phosphate groups (as may typically be found in RNA or DNA), or modified or substituted sugar or phosphate groups. Alternatively, the backbone of the polynucleotide can comprise a polymer of synthetic subunits such as phosphoramidites and thus can be an oligodeoxynucleoside phosphoramidate or a mixed phosphoramidate-phosphodiester oligomer.

Peyrottes et al. (1996) Nucl. Acids Res. 24:1841-1848; Chaturvedi et al. (1996) Nucl. Acids Res. 24:2318-2323. A polynucleotide may comprise modified nucleotides, such as methylated nucleotides and nucleotide analogs, uracyl, other sugars, and linking groups such as fluororibose and thioate, and nucleotide branches. The sequence of nucleotides may be interrupted by non-nucleotide components. A polynucleotide may be further modified after polymerization, such as by conjugation with a labeling component. Other types of modifications included in this definition are caps, substitution of one or more of the naturally occurring nucleotides with an analog, and introduction of means for attaching the polynucleotide to proteins, metal ions, labeling components, other polynucleotides, or a solid support.

The terms "polypeptide" and "protein", used interchangeably herein, refer to a polymeric form of amino acids of any length, which can include coded and non-coded amino acids, chemically or biochemically modified or derivatized amino acids, and polypeptides having modified peptide backbones. The term includes fusion proteins, including, but not limited to, fusion proteins with a heterologous amino acid sequence, fusions with heterologous and homologous leader sequences, with or without N-terminal methionine residues; immunologically tagged proteins; and the like.

A polynucleotide or polypeptide has a certain percent "sequence identity" to another polynucleotide or polypeptide, meaning that, when aligned, that percentage of bases or amino acids are the same when comparing the two sequences. Sequence similarity can be determined in a number of different manners. To determine sequence identity, sequences can be aligned using the methods and computer programs, including BLAST, available over the world wide web at

<http://www.ncbi.nlm.nih.gov/BLAST/>. Another alignment algorithm is FASTA, available in the Genetics Computing Group (GCG) package, from Madison, Wisconsin, USA, a wholly owned subsidiary of Oxford Molecular Group, Inc. Other techniques for alignment are described in Methods in Enzymology, vol. 266: Computer Methods for Macromolecular Sequence Analysis (1996), ed.

5 Doolittle, Academic Press, Inc., a division of Harcourt Brace & Co., San Diego, California, USA. Of particular interest are alignment programs that permit gaps in the sequence. The Smith-Waterman is one type of algorithm that permits gaps in sequence alignments. See Meth. Mol. Biol. 70: 173-187 (1997). Also, the GAP program using the Needleman and Wunsch alignment method can be utilized to align sequences. See J. Mol. Biol. 48: 443-453 (1970)

10 Of interest is the BestFit program using the local homology algorithm of Smith Waterman (Advances in Applied Mathematics 2: 482-489 (1981) to determine sequence identity. The gap generation penalty will generally range from 1 to 5, usually 2 to 4 and in many embodiments will be 3. The gap extension penalty will generally range from about 0.01 to 0.20 and in many instances will be 0.10. The program has default parameters determined by the sequences inputted to be compared.

15 Preferably, the sequence identity is determined using the parameters determined by the program. This program is available also from Genetics Computing Group (GCG) package, from Madison, Wisconsin, USA.

Another program of interest is the FastDB algorithm. FastDB is described in Current Methods in Sequence Comparison and Analysis, Macromolecule Sequencing and Synthesis, Selected 20 Methods and Applications, pp. 127-149, 1988, Alan R. Liss, Inc. Percent sequence identity is calculated by FastDB based upon the following parameters:

Mismatch Penalty: 1.00;  
Gap Penalty: 1.00;  
Gap Size Penalty: 0.33; and  
25 Joining Penalty: 30.0.

One parameter for determining percent sequence identity is the "percentage of the alignment region length" where the strongest alignment is found.

The percentage of the alignment region length is calculated by counting the number of residues of the individual sequence found in the region of strongest alignment. This number is divided by the 30 total residue length of the target or query polynucleotide sequence to find a percentage. An example is shown below:

Target sequence: GCGCGAAATACTCACTCGAGG  
                          |      |||  ||||  |||  
Query sequence: TATAGCCCTAC . CACTAGAGTCC

35                   1     5     10     15

The region of alignment begins at residue 9 and ends at residue 19. The total length of the target sequence is 20 residues. The percent of the alignment region length is 11 divided by 20 or 55%, for example.

Percent sequence identity is calculated by counting the number of residue matches between the 5 target and query polynucleotide sequence and dividing total number of matches by the number of residues of the target or query sequence found in the region of strongest alignment. For the example above, the percent identity would be 10 matches divided by 11 residues, or approximately, 90.9%.

The percent of the alignment region length is typically at least about 55% of total length of the sequence, more typically at least about 58%, and even more typically at least about 60% of the total 10 residue length of the sequence. Usually, percent length of the alignment region can be as great as about 62%, more usually as great as about 64% and even more usually as great as about 66%.

"Recombinant," as used herein, means that a particular DNA sequence is the product of various combinations of cloning, restriction, and ligation steps resulting in a construct having a structural coding sequence distinguishable from homologous sequences found in natural systems. 15 Generally, DNA sequences encoding the structural coding sequence, for example cytokines, can be assembled from cDNA fragments and short oligonucleotide linkers, or from a series of oligonucleotides, to provide a synthetic gene which is capable of being expressed in a recombinant transcriptional unit. Such sequences are preferably provided in the form of an open reading frame uninterrupted by internal nontranslated sequences, or introns, which are typically present in eukaryotic 20 genes. Genomic DNA containing the relevant sequences could also be used. Sequences of non-translated DNA may be present 5' or 3' from the open reading frame, where such sequences do not interfere with manipulation or expression of the coding regions. Thus, the term "recombinant" polynucleotide or nucleic acid refers to one which is not naturally occurring, or is made by the artificial combination of two otherwise separated segments of sequence. This artificial combination is 25 often accomplished by either chemical synthesis means, or by the artificial manipulation of isolated segments of nucleic acids, e.g., by genetic engineering techniques. Such is usually done to replace a codon with a redundant codon encoding the same or a conservative amino acid, while typically introducing or removing a sequence recognition site. Alternatively, it is performed to join together nucleic acid segments of desired functions to generate a desired combination of functions.

30 By "antisense polynucleotide" is mean a polynucleotide having a nucleotide sequence complementary to a given polynucleotide sequence (e.g., a polynucleotide sequence encoding a soluble adenylyl cyclase polypeptide) including polynucleotide sequences associated with the transcription or translation of the given polynucleotide sequence (e.g., a promoter of a polynucleotide encoding sAC polypeptide), where the antisense polynucleotide is capable of hybridizing to a human sAC 35 polypeptide-encoding polynucleotide sequence. Of particular interest are antisense polynucleotides

capable of inhibiting transcription and/or translation of a sAC-encoding polynucleotide either in vitro or in vivo.

"Peptide nucleic acid" as used herein refers to a molecule which comprises an oligomer to which an amino acid residue, such as lysine, and an amino group have been added. These small molecules, also designated anti-gene agents, stop transcript elongation by binding to their complementary (template) strand of nucleic acid (Nielsen et al. (1993) *Anticancer Drug Des.* 8:53-63).

The term "specific binding" with respect to antibody-antigen interactions, is a term well understood in the art and indicates that a given antibody binds to an antigenic determinant (epitope) which was used as an immunogen, and does not substantially bind to an unrelated antigenic determinant.

As used herein the term "isolated" is meant to describe a polynucleotide, a polypeptide, an antibody, or a host cell that is in an environment different from that in which the polynucleotide, the polypeptide, the antibody, or the host cell naturally occurs. As used herein, the term "substantially purified" refers to a compound (e.g., either a polynucleotide or a polypeptide or an antibody) that is removed from its natural environment and is at least 60% free, preferably 75% free, and most preferably 90% free from other components with which it is naturally associated.

The term "host cell" includes an individual cell or cell culture which can be or has been a recipient of any recombinant vector(s) or isolated polynucleotide of the invention. Host cells include progeny of a single host cell, and the progeny may not necessarily be completely identical (in morphology or in total DNA complement) to the original parent cell due to natural, accidental, or deliberate mutation and/or change. A host cell includes cells transfected or infected *in vivo* or *in vitro* with a recombinant vector or a polynucleotide of the invention. A host cell which comprises a recombinant vector of the invention is a "recombinant host cell".

The term "immunologically active" defines the capability of the natural, recombinant or synthetic human sAC polypeptide, or any oligopeptide thereof, to induce a specific immune response in appropriate animals or cells and to bind with specific antibodies. As used herein, "antigenic amino acid sequence" means an amino acid sequence that, either alone or in association with a carrier molecule, can elicit an antibody response in a mammal.

By "transformation" is meant a permanent or transient genetic change induced in a cell following incorporation of new DNA (i.e., DNA exogenous to the cell). Genetic change can be accomplished either by incorporation of the new DNA into the genome of the host cell, or by transient or stable maintenance of the new DNA as an episomal element. Where the cell is a mammalian cell, a permanent genetic change is generally achieved by introduction of the DNA into the genome of the cell.

By "construct" is meant a recombinant nucleic acid, generally recombinant DNA, that has been generated for the purpose of the expression of a specific nucleotide sequence(s), or is to be used in the construction of other recombinant nucleotide sequences.

5 A "transcriptional control region" (sometimes referred to as a "transcriptional regulatory region") encompasses all the elements necessary for transcription, and may include elements necessary for transcription. Thus, a transcriptional control region includes at least the promoter sequence, and may also include other regulatory sequences such as enhancers, and transcription factor binding sites.

A "transcriptional control region heterologous to a coding region" is one that is not normally associated with the coding region in nature.

10 "Operably linked" refers to a juxtaposition wherein the components so described are in a relationship permitting them to function in their intended manner. For instance, a promoter is operably linked to a coding sequence if the promoter effects its transcription or expression.

15 "Regulatory sequences" refer to those sequences normally associated with (for example within 50 kb) of the coding region of a locus which affect the expression of the gene (including transcription of the gene, and translation, splicing, stability, or the like of the messenger RNA). Regulatory sequences include, *inter alia*, promoters, enhancers, splice sites and polyadenylation sites.

20 The term "biological sample" encompasses a variety of sample types obtained from an organism and can be used in a diagnostic or monitoring assay. The term encompasses blood and other liquid samples of biological origin, solid tissue samples, such as a biopsy specimen or tissue cultures or cells derived therefrom and the progeny thereof. The term encompasses samples that have been manipulated in any way after their procurement, such as by treatment with reagents, solubilization, or enrichment for certain components. The term encompasses a clinical sample, and also includes cells in cell culture, cell supernatants, cell lysates, serum, plasma, biological fluids, and tissue samples.

25 By "individual" or "subject" or "patient" is meant any mammalian subject for whom diagnosis, treatment, or therapy is desired, particularly humans, particularly male humans. Other subjects may include cattle, dogs, cats, guinea pigs, rabbits, rats, mice, horses, and so on.

#### METHODS OF THE INVENTION

The present invention provides methods for reducing the number of motile sperm in a male. 30 The invention further provides methods for inhibiting or reducing spermatogenesis in a male subject, and for reducing sperm motility in a male subject. Such methods are useful in male contraception. The male may be a human male, or may be a non-human male, including, for example, a male in a non-human animal population, such as an undesired rodent population.

The present invention also provides methods for identifying agents which modulate a level of sAC enzyme activity; methods for identifying agents which modulate a level of sAC mRNA; methods for identifying agents which modulate a level of sAC polypeptide.

The present invention further provides methods for detecting the presence and/or measuring an 5 amount of a human sAC mRNA in a biological sample; and methods of detecting the presence of and/or measuring an amount of human sAC polypeptide in a biological sample.

#### Methods of reducing the number of motile sperm

The present invention provides methods of reducing the number of motile sperm in a male.

10 Reduction of the number of motile sperm can occur by inhibiting spermatogenesis, and/or by inhibiting sperm motility, in a male. The methods generally comprise administering to the individual a composition comprising an effective amount of a substance that inhibits sAC adenylyl cyclase activity, or that inhibits production of sAC mRNA and/or sAC polypeptide, i.e., administration of the substance results in reduced levels of active sAC. These methods are useful in achieving male contraception. A 15 reduction in numbers of motile sperm may be an indication that the substance reduces spermatogenesis, or that the substance reduces sperm motility. The invention further provides methods for reducing sperm motility in a male. The invention further provides methods for reducing or inhibiting spermatogenesis in a male.

Substances of interest which inhibit sAC adenylyl cyclase are those that inhibit (reduce) 20 spermatogenesis, e.g., that reduce the number of sperm produced by a male, particularly the number of motile sperm produced by a male; and substances that inhibit sperm motility. Inhibition of spermatogenesis can be determined by determining the number of motile and non-motile sperm in a sperm sample. Generally, an "effective amount" of a substance is an amount that is effective in reducing the number of motile sperm produced by the male to which the substance is administered. An 25 effective amount of a substance results in a reduction of motile sperm production, as measured in a sperm sample from the subject, to less than about 10 million spermatozoa/mL, typically less than about 5 million spermatozoa/mL, usually less than about 1 million spermatozoa/mL. World Health Organization standards can also be used to assess the effect of a substance on fertility. The WHO standard for oligospermia is  $20 \times 10^6$  sperm/ml, and for severe oligospermia is  $3-5 \times 10^6$  sperm/ml, 30 with fewer than 50% motility in each case. Motility can be determined by any known means, including, but not limited to, CASA, or computer-assisted sperm analysis, which measures various sperm motility parameters over time.

The invention further provides methods of male contraception. In some embodiments, the methods are reversible, e.g., when the substance is no longer administered to the individual, the level of

fertility (i.e., motile sperm production) which existed in the individual before administration of the substance is restored to the individual, usually within a few days, or weeks, up to about a few months. The methods generally comprise administering to a male a pharmaceutical composition comprising an effective amount of a substance which preferentially inhibits soluble adenylyl cyclase, or which 5 preferentially reduces sAC mRNA synthesis and/or sAC polypeptide synthesis. A substance which preferentially inhibits soluble adenylyl cyclase, or which preferentially reduces sAC mRNA synthesis and/or sAC polypeptide synthesis, is one that inhibits the testis-specific soluble adenylyl cyclase, or which reduces testis-specific sAC mRNA synthesis and/or testis specific sAC polypeptide synthesis, as described herein, but which does not substantially reduce activity of other, non-sAC, enzymes, or 10 which does not substantially reduce synthesis of other, non-sAC, mRNA species or synthesis of other, non-sAC, proteins. Thus, for example, transmembrane, G protein-coupled, adenylyl cyclase activity and/or synthesis is substantially not affected by the substance, e.g., the activity and/or synthesis of a transmembrane, G protein-coupled adenylyl cyclase retains at least about 90% of its activity and/or synthesis in the presence of the substance.

15 The methods can be applied to a wide variety of species, including mammals, reptiles, amphibians, insects, arachnids, etc. Of particular interest are methods of reducing sperm production (e.g., reducing the number of motile sperm) in human males. Also of interest are methods of reducing sperm production in unwanted rodent populations. Also of interest are methods of reducing sperm production in feral mammalian populations (e.g., feral cats, feral dogs), where it is desired to reduce 20 the number of offspring of such animals. Of further interest are methods of reducing spermatogenesis in unwanted insect populations, including, but not limited to, mosquitoes, fleas, weevils, locusts, and the like.

25 Reduction of sperm production can be species-specific, e.g., the substance administered can be one that inhibits sperm production only in human males. Alternatively, the substance can be one that reduces sperm production in more than one species, e.g., of interest are substances that inhibit sperm production in both rats and mice.

Whether spermatogenesis has been inhibited or reduced can be determined by any known method. For example, a sperm sample can be obtained from the individual at a suitable time after the composition has been administered to the individual. The sperm sample can be a freshly collected 30 ejaculate; seminal plasma-free sperm, e.g., sperm that has been substantially isolated from other semen components, such as by centrifugation. The total number of motile and non-motile sperm is counted. Sperm can be quantitated based on detecting a component of sperm, or can be performed visually, e.g., counting the number of sperm visually under a microscope.

35 For determining whether a particular male contraceptive has been effective, the test must be particularly sensitive. For example, the VasScore™ test, can successfully distinguish colors based on a

quantity of sperm protein that corresponds to a sperm concentration in the sample that is greater than or equal to about 100,000 spermatozoa/mL. If the sperm sample is semen, it is preferred that the semen is liquefied prior to analysis. Although a semen sample will typically liquefy on its own at room temperature in about 30-60 minutes, preferably the semen sample is liquefied in less than 30 minutes and most preferably it is liquefied in 5-15 minutes, based on contact with a liquefaction reagent.

5 Examples of appropriate liquefaction reagents include non-enzymatic reagents, such as dithiothreitol (DTT) (e.g. 1-5 mg/ml) and enzymes such as chymotrypsin and pronase (e.g. 5-15 mg/ml.)

Methods for determining sperm fertility potential include those involving quantitating a particular sperm target or component as an indication of the number of sperm present in a sperm 10 containing sample. Examples of appropriate targets or components include sperm proteins (e.g. sperm flagellar proteins, glycolytic enzymes, antioxidant enzymes (e.g. glutathione peroxidase or superoxide dismutase), nuclear proteins, acrosomal proteins,  $\alpha$ -tubulin, lactate dehydrogenase (LDH-X), protamine (sperm histones), acrosomal proteins (e.g. acrosin) or mitochondrial proteins); sperm lipids (e.g. cholesterol, phospholipids, glycolipids, triglycerides, phosphatidylglycerols, 15 seminolipids, and fatty acids, particularly docosahexaenoic acid, which is one of the few fatty acids found in sperm); nucleic acids or a mixture of sperm components (e.g. thiazine blue reacts with sperm proteins, lipids and other sperm components).

An example of a test for male fertility which can be used in the present invention is described in U.S. Patent No. 5,935,800.

20 Suitable sperm reagents include labeled (e.g. dye or tracer labeled) or unlabeled reagents that specifically stain based on interaction with a sperm component (e.g. sperm protein, sperm lipid, sperm nucleic acid, sperm carbohydrate and/or other sperm component (e.g. mitochondria, nucleus). For example, Protein Reagent (0.3% tetrabromophenol, Miles Scientific, Connecticut) colors sperm based on interaction with sperm protein. Rhodamine 123 colors sperm by accumulating within sperm 25 mitochondria. Detectably labeled protein A specifically colors sperm by binding to sperm bound antibodies. Propidium iodide and eosin specifically color sperm by interacting with sperm nucleic acids. However, both reagents can only diffuse into cells which have been permeabilized (e.g. using a detergent). Acridine orange, on the other hand, can specifically color unpermeabilized sperm based on interaction with decondensed sperm chromatin (but not condensed sperm chromatin). Sperm chromatin 30 can be decondensed prior to staining with acridine orange by reducing sperm protamines (i.e. sperm histones). A preferred reagent for reducing sperm protamines is dithiothreitol (DTT). Other reagents, which specifically interact with a semen or washed sperm sample, include gold particles (which have an intrinsic pink color and react with proteins) thiazine blue, tetrabromophenol and rhodamine (red-colored and reacts with lipids). Other preferred sperm reagents include sperm antibodies, such as 35 labeled (e.g. enzyme, tracer (e.g. radioactive), dye or color particle labeled) or unlabeled anti-sperm

antibodies (e.g. anti-human sperm polyclonal antibody; Arnel Products Co., Inc, Cherokee Station, New York, N.Y.; Chemicon International Inc., Temecula, Calif.) or labeled or unlabeled antibodies against a sperm component (e.g. a sperm protein or sperm lipid). Preferred antibodies include anti-human sperm polyclonal antibodies and antibodies specific to an epitope of the sperm flagellum, 5 nuclear proteins, glycolytic enzymes, acrosome etc.).

One method for quantitating sperm involves (i) incubating a sperm sample with colored particles containing anti-sperm antibodies for an appropriate period of time to allow the sperm antigens to react with the antibody bound colored particles; ii) filtering the sample of step i), so that sperm/colored particle/antibody complex is retained on the filter and unbound, colored 10 particle/antibody and seminal plasma protein passes through the filter; and iii) visualizing the color intensity on the filter as an indication of the number of sperm on the filter. For example, if colored particles are used, sperm/colored particle/antibody complexes can be quantitated by comparing the color of the filter to a color chart, which depicts various color possibilities for various quantities of sperm.

15 In order to use antibodies or reagents that may also react with seminal plasma components present in a sperm containing sample (e.g. ejaculate), sperm can first be isolated. Seminal plasma-free sperm can then be contacted with anti-sperm antibody coated colored particles. After a sufficient period of time to allow antibodies and antigens to react, unbound antibody coated colored particles can be removed from the mixture and sperm/colored particle/antibody complex detected and quantitated. 20 Alternatively, sperm can be quantitated by detecting the appearance of agglutination in a drop of sample following addition of anti-sperm antibodies with bound latex particles.

25 Immunodetection of an antigenic indicator of sperm in a sample can be accomplished using any of a number of competitive or non-competitive assay procedures. In general, competitive immunoassays are performed by adding the antigen to be detected to a sperm containing sample, so that the sperm and the antigen compete for a limited number of antibody binding sites resulting in the formation of sperm-antibody and labeled antigen-antibody complexes. By maintaining the concentration of labeled antigen and antibody constant, the amount of labeled antibody complex formed is inversely proportional to the amount of sperm present in the sample. A quantitative determination of the sperm can therefore be made based on the labeled antibody complex.

30 Competitive assays can be homogeneous (i.e. not requiring separation of antibody bound tracer (e.g. labeled antigen) from free tracer, since the antigen-antibody interaction causes, directly or indirectly, a measurable change in the signal obtained from the label group of the tracer). Alternatively, competitive assays can be heterogeneous (i.e. requiring separation of bound tracer from free tracer prior to determining the amount of ligand in the sample).

In contrast to competitive immunoassays, non-competitive assays involve incubating a sperm containing sample with an immobilized sperm antibody for a period of time sufficient to reach equilibrium with regard to the formation of antibody-sperm conjugates. The sperm antibody can be directly or indirectly labeled. For example, indirect labeling can be carried out after a 5 wash step to remove unbound sperm by contacting the immobilized antibody-sperm complexes with a second, labeled antibody that is specific for the antibody-sperm complex. Following a second wash step to remove unbound second antibody, the amount of bound second antibody can be detected and measured as an indication of bound sperm.

10 Methods of identifying substances that modulate sAC enzyme activity

The present invention provides methods of identifying agents which modulate an adenylyl cyclase activity of an sAC polypeptide. In some embodiments, the methods are cell-based methods. In other embodiments, the methods are cell-free methods. The term "modulate" encompasses an increase or a decrease in the measured adenylyl cyclase activity when compared to a suitable control.

15 The methods generally comprise:

- a) contacting a substance to be tested with a sample containing an sAC polypeptide; and
- b) assaying an adenylyl cyclase activity of the human sAC polypeptide in the presence of the substance, wherein an increase or a decrease in adenylyl cyclase activity in comparison to sAC 20 adenylyl cyclase activity in a suitable control is an indication that the substance modulates an adenylyl cyclase activity of the sAC.

The sAC polypeptide used in these methods is one that is preferentially produced in testis, in particular, in spermatids, of the species from which it was derived. sAC polypeptides are produced preferentially in those cells which preferentially express sAC mRNA. The term "preferentially expressed in spermatids" indicates that sAC mRNA is expressed at a level generally at least about 2-fold, usually at least about 10-fold, typically at least about 20-fold or more higher in spermatids than 25 in somatic cells of the body. A convenient reference cell is the Sertoli cell, which does not express sAC mRNA. Whether an sAC mRNA is expressed preferentially in spermatids can be readily determined by those skilled in the art, by determining the level of sAC mRNA in spermatids, in comparison with sAC mRNA levels in a somatic cell, including, but not limited to, Sertoli cells. 30 Methods of measuring mRNA levels are known in the art, and any of these methods are suitable for use herein. Suitable methods include Northern analysis, and PCR.

The sAC polypeptide can be a full-length polypeptide (e.g., has an amino acid sequence of the same length as that found in its natural environment, or "wild-type" sequence), but need not be full-length, as long as the sAC polypeptide retains measurable adenylyl cyclase activity. Furthermore, the 35 sAC polypeptide used in these assays may also contain alterations in amino acid sequence compared to

the wild-type sequence, wherein such alterations may confer a desirable property, including, but not limited to, enhanced stability *in vitro*, and the like. The sAC polypeptide may further be a fusion protein comprising an sAC polypeptide and a heterologous polypeptide, e.g. a non-sAC polypeptide, including, but not limited to, an epitope to facilitate recovery of the sAC from the sample, and the like.

5 In some embodiments, the sAC polypeptide is a human sAC polypeptide of the invention. In other embodiments, the sAC polypeptide is one derived from non-human species, including, but not limited to, a non-human mammalian species, such as rat. For example, polypeptides comprising the sequences set forth in SEQ ID NO:2, which depicts human sAC, and SEQ ID NO:4, which depicts a rat sAC amino acid sequence, or fragments of these sequences which retain adenylyl cyclase activity, 10 can be used.

Where the method is a cell-free assay method, sAC enzyme may be, but need not be, substantially purified. In general, the enzyme should be isolated from the source of the enzyme in those instances where one or more components found in the source of the enzyme interfere with enzyme activity or measurement of enzyme activity. The sample can be a cell lysate comprising sAC, 15 or the sample can comprise sAC which is purified to any degree. As non-limiting examples, the sample can be: a cell lysate of a mammalian cell line which has been transfected with a recombinant vector ("construct") which encodes and expresses sAC polypeptide having adenylyl cyclase activity; and sAC which has been purified from a biological source.

sAC adenylyl cyclase activity can be measured in a cell-free system (e.g., in cell lysates, or in 20 a sample comprising isolated sAC), using any known assay, such as those described in Buck et al. (1999) *Proc. Natl. Acad. Sci. USA* 96:79-84; Steiner et al. (1974) *Methods Enzymol.* 38:96-105; and Harper et al. (1975) *J. Cyclic Nucleotide Res.* 1:207-218. sAC activity is generally measured by measuring an amount of cAMP produced. sAC activity may be measured in the presence of labeled substrate ATP, thereby generating labeled product cAMP. For example, [ $\alpha$ -<sup>32</sup>P]ATP, or any other 25 labeled form of ATP can be used, as long as the product cAMP is detectably labeled. Typically, these assays are conducted in the presence of 5 mM MnCl<sub>2</sub> and 5 mM [ $\alpha$ -<sup>32</sup>P]ATP. sAC activity may also be measured by measuring a level of cAMP using cAMP-specific antibodies, using, e.g., an ELISA or an RIA. Commercially available kits may also be used.

The following is a non-limiting example of how sAC activity can be measured in cell lysates. 30 Mammalian cells are transfected with a recombinant vector comprising a nucleotide sequence which encodes an sAC. After two days, cells are labeled overnight with 5 mM [ $\alpha$ -<sup>32</sup>P]ATP (specific activity 4 x 10<sup>4</sup> cpm/nmol). The next day, cells are suspended in 200  $\mu$ l of lysis buffer (50 mM Tris (pH 7.5), 1 mM EDTA, 1 mM dithiotreitol, 0.1 mg/ml leupeptin, 1 mM phenylmethylsulfonyl fluoride) and disrupted by sonication for 1 minute on ice. Extracts are centrifuged at 100,000 x g for 10 minutes. 35 The supernatant, i.e., cytosolic fraction, is assayed for sAC activity in the presence of 5 mM MnCl<sub>2</sub>.

Where the method is a cell-based assay method, a cell which synthesizes sAC is contacted with the compound being tested, and, after a suitable time, the cells are lysed, and sAC activity is measured in the cell lysates. Alternatively, the cells need not be lysed in order to measure sAC adenylyl cyclase activity. In these embodiments, cAMP levels are measured in intact cells. As a non-limiting example, a construct comprising nucleotide sequence encoding an sAC polypeptide is introduced into a cell line, e.g., COS cells, HEK293 cells, or MA-10 cells, such that sAC is expressed in the cells. For these assays, the sAC coding region may be under control of an endogenous promoter, or, alternatively, under control of an inducible promoter. Inducible promoters are known in the art, and can be used in such a construct. Suitable inducible promoters include, but are not limited to, a hormone-inducible promoter. When an inducible promoter is used, the inducer is added to the cell culture before, or simultaneously with, the substance being tested. Controls include a culture to which no inducer has been added, as well as a culture to which inducer, but no substance being tested, is added.

Alternatively, endogenous sAC activity can be measured in intact cells.

sAC adenylyl cyclase activity can be measured in an intact cell using any known assay, such as those described above, wherein accumulation of labeled cAMP is measured in intact cells. A non-limiting example of such an assay is a fluorescence energy transfer assay, as described in U.S. Patent No. 5,439,797. Commercially available kits may also be used.

Assays such as those described herein are amenable to high through-put screening assays.

For example, isolated sAC, or cells comprising endogenous sAC, or cells expressing a construct encoding sAC, each in separate well of a microtiter plate, e.g., can be contacted with a large number of test compounds at a time, thereby allowing automation.

The term "agent" is used interchangeably herein with the terms "substance" and "compound". An "agent which modulates an adenylyl cyclase activity of a sAC polypeptide", as used herein, describes any molecule, e.g. protein; peptide; natural or synthetic inorganic or organic compound, or pharmaceutical, with the capability of altering an adenylyl cyclase activity of a sAC polypeptide, as described herein. Generally a plurality of assay mixtures are run in parallel with different agent concentrations to obtain a differential response to the various concentrations. Typically, one of these concentrations serves as a negative control, i.e. at zero concentration or below the level of detection.

Candidate agents encompass numerous chemical classes, and may be natural or synthetic inorganic or organic molecules, which may be small organic compounds having a molecular weight of more than 50 and less than about 2,500 daltons. Candidate agents may comprise functional groups necessary for structural interaction with proteins, particularly hydrogen bonding, and typically include at least an amine, carbonyl, hydroxyl or carboxyl group, preferably at least two of the functional chemical groups. The candidate agents often comprise cyclical carbon or heterocyclic structures

and/or aromatic or polycyclic structures substituted with one or more of the above functional groups. Candidate agents are also found among biomolecules including peptides, saccharides, fatty acids, steroids, purines, pyrimidines, derivatives, structural analogs or combinations thereof.

Candidate agents are obtained from a wide variety of sources including libraries of synthetic or natural compounds. For example, numerous means are available for random and directed synthesis of a wide variety of organic compounds and biomolecules, including expression of randomized oligonucleotides and oligopeptides. Alternatively, libraries of natural compounds in the form of bacterial, fungal, plant and animal extracts are available or readily produced. Additionally, natural or synthetically produced libraries and compounds are readily modified through conventional chemical, physical and biochemical means, and may be used to produce combinatorial libraries. Known pharmacological agents may be subjected to directed or random chemical modifications, such as acylation, alkylation, esterification, glycosylation, amidification, etc. to produce structural analogs.

A variety of other reagents may be included in the screening assay. These include reagents like salts, neutral proteins, e.g. albumin, detergents, etc that are used to facilitate optimal protein-protein binding and/or reduce non-specific or background interactions. Reagents that improve the efficiency of the assay, such as protease inhibitors, nuclease inhibitors, anti-microbial agents, etc. may be used. The mixture of components are added in any order that provides for the requisite binding. Incubations are performed at any suitable temperature, typically between 4 and 40 °C. Incubation periods are selected for optimum activity, but may also be optimized to facilitate rapid high-throughput screening. Typically between 0.1 and 1 hours will be sufficient.

Adenylyl cyclase activity can be measured using any adenylyl cyclase assay known in the art. An agent which modulates an adenylyl cyclase activity of a human sAC polypeptide increases or decreases the activity at least about 10%, more preferably at least about 25%, more preferably at least about 50%, more preferably at least about 100%, or 2-fold, more preferably at least about 5-fold, more preferably at least about 10-fold or more when compared to a suitable control.

Agents which increase or decrease an adenylyl cyclase activity of a human sAC polypeptide to the desired extent may be selected for further study, and assessed for bioavailability, cellular availability, cytotoxicity, biocompatibility, etc.

Agents which decrease an adenylyl cyclase activity of a human sAC polypeptide may find use in methods for inhibiting spermatogenesis, and thus may find use as male contraceptives.

#### Methods of detecting agents which modulate a level of sAC mRNA and/or sAC polypeptide

A wide variety of cell-based assays may be used for identifying agents which modulate levels of human sAC mRNA, using, for example, a mammalian cell transformed with a construct comprising

a human sAC-encoding cDNA such that the cDNA is overexpressed. In these assays, a level of sAC mRNA or polypeptide is measured.

Alternatively, a construct comprising a cAMP-sensitive promoter operably linked to a reporter gene, e.g., luciferase, can be used. In such, a level of sAC produced is measured in the presence and absence of a substance being tested. Inhibition of sAC mRNA and/or polypeptide production is measured in terms of a decrease in the level of signal produced by the reporter gene product. An example of such a construct is one which comprises a cAMP-responsive element (CRE) operably linked to reporter gene encoding luciferase. A multiplicity of CRE can be used, e.g., three CRE in tandem.

Accordingly, the present invention provides a method for identifying an agent, particularly a biologically active agent, that modulates a level of human sAC expression in a cell, the method comprising: combining a candidate agent to be tested with a cell comprising a nucleic acid which encodes a human sAC polypeptide; and determining the effect of said agent on human sAC expression. "Modulation" of human sAC expression levels includes increasing the level and decreasing the level of human sAC mRNA and/or human sAC polypeptide encoded by the human sAC polynucleotide when compared to a control lacking the agent being tested. An increase or decrease of about 1.25-fold, usually at least about 1.5-fold, usually at least about 2-fold, usually at least about 5-fold, usually at least about 10-fold or more, in the level (i.e., an amount) of human sAC mRNA and/or polypeptide following contacting the cell with a candidate agent being tested, compared to a control to which no agent is added, is an indication that the agent modulates human sAC expression.

An agent being tested for its effect on human sAC expression is assessed for any cytotoxic activity it may exhibit toward the cell used in the assay, using well-known assays, including, but not limited to, trypan blue dye exclusion, an MTT ([3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl-2 H-tetrazolium bromide]) assay, and the like. Agents that do not exhibit cytotoxic activity are considered candidate agents.

The cells used in the assay are usually mammalian cells, including, but not limited to, rodent cells and human cells. The cells may be primary cultures of spermatids, or may be immortalized cell lines, e.g., COS cells, MA-10 cells, HEK293 cells, and the like.

Human sAC mRNA and/or polypeptide whose levels are being measured can be encoded by an endogenous human sAC polynucleotide, or the human sAC polynucleotide can be one that is comprised within a recombinant vector and introduced into the cell, i.e., the human sAC mRNA and/or polypeptide can be encoded by an exogenous human sAC polynucleotide. For example, a recombinant vector may comprise an isolated human sAC transcriptional regulatory sequence, such as a promoter sequence, operably linked to a reporter gene (e.g.,  $\beta$ -galactosidase, CAT, luciferase, or other gene that can be easily assayed for expression). In these embodiments, the method for identifying an

agent that modulates a level of human human sAC expression in a cell, comprises: combining a candidate agent to be tested with a cell comprising a nucleic acid which comprises a human sAC gene transcriptional regulatory element operably linked to a reporter gene; and determining the effect of said agent on reporter gene expression. A recombinant vector may comprise an isolated human sAC transcriptional regulatory sequence, such as a promoter sequence, operably linked to sequences coding for a human sAC polypeptide; or the transcriptional control sequences can be operably linked to coding sequences for a human sAC fusion protein comprising human sAC polypeptide fused to a polypeptide which facilitates detection. In these embodiments, the method comprises combining a candidate agent to be tested with a cell comprising a nucleic acid which comprises a human sAC gene transcriptional regulatory element operably linked to a human sAC polypeptide-coding sequence; and determining the effect of said agent on human sAC expression, which determination can be carried out by measuring an amount of human sAC mRNA, human sAC polypeptide, or human sAC fusion polypeptide produced by the cell.

Cell-based assays generally comprise the steps of contacting the cell with an agent to be tested, forming a test sample, and, after a suitable time, assessing the effect of the agent on human sAC expression. A control sample comprises the same cell without the candidate agent added. human sAC expression levels are measured in both the test sample and the control sample. A comparison is made between human sAC expression level in the test sample and the control sample. human sAC expression can be assessed using conventional assays. For example, when a mammalian cell line is transformed with a construct that results in expression of human sAC, human sAC mRNA levels can be detected and measured, as described above, or human sAC polypeptide levels can be detected and measured, as described above. A suitable period of time for contacting the agent with the cell can be determined empirically, and is generally a time sufficient to allow entry of the agent into the cell and to allow the agent to have a measurable effect on human sAC mRNA and/or polypeptide levels.

Generally, a suitable time is between 10 minutes and 24 hours, more typically about 1-8 hours. Methods of measuring human sAC mRNA levels are known in the art, several of which have been described above, and any of these methods can be used in the methods of the present invention to identify an agent which modulates human sAC mRNA level in a cell, including, but not limited to, a PCR, such as a PCR employing detectably labeled oligonucleotide primers, and any of a variety of hybridization assays. The use of the polymerase chain reaction is described in Saiki *et al.* (1985) *Science* 239:487, and a review of current techniques may be found in Sambrook, *et al.* Molecular Cloning: A Laboratory Manual, CSH Press 1989, pp.14.2-14.33. A detectable label may be included in the amplification reaction. Suitable labels include fluorochromes, *e.g.* fluorescein isothiocyanate (FITC), rhodamine, Texas Red, phycoerythrin, allophycocyanin, 6-carboxyfluorescein (6-FAM), 2',7'-dimethoxy-4',5'-dichloro-6-carboxyfluorescein (JOE), 6-carboxy-X-rhodamine (ROX),

6-carboxy-2',4',7',4,7-hexachlorofluorescein (HEX), 5-carboxyfluorescein (5-FAM) or N,N,N',N'-tetramethyl-6-carboxyrhodamine (TAMRA), radioactive labels, *e.g.*  $^{32}\text{P}$ ,  $^{35}\text{S}$ ,  $^{3}\text{H}$ ; *etc.* The label may be a two stage system, where the amplified DNA is conjugated to biotin, haptens, *etc.* having a high affinity binding partner, *e.g.* avidin, specific antibodies, *etc.*, where the binding partner is conjugated to a detectable label. The label may be conjugated to one or both of the primers. Alternatively, the pool of nucleotides used in the amplification is labeled, so as to incorporate the label into the amplification product.

Similarly, human sAC polypeptide levels can be measured using any standard method, several of which have been described herein, including, but not limited to, an immunoassay such as ELISA, 10 for example an ELISA employing a detectably labeled antibody specific for a human sAC polypeptide.

The method described above is useful for identifying agents which may be useful in reducing spermatogenesis. An agent which reduces human sAC gene expression and is not cytotoxic is considered a possible agent for male contraceptive. Such agents are then further evaluated for safety and efficacy.

15

#### Screening of Candidate Agents Using Cell-Free Assays

Cell-free assays, *i.e.*, assays which measure human sAC polypeptide levels or function directly, include, but are not limited to, adenylyl cyclase activity assays, labeled *in vitro* protein-protein binding assays, protein-DNA binding assays, electrophoretic mobility shift assays, 20 immunoassays for protein binding, and the like. Using these methods, one can identify substances that bind specifically to human sAC polypeptides. Such substances are useful as diagnostic agents to detect the presence of and/or to measure a level of human sAC polypeptide in a biological.

The screening assay can be a binding assay, wherein one or more of the molecules may be joined to a label, and the label directly or indirectly provide a detectable signal. Various labels include 25 radioisotopes, fluorescers, chemiluminescers, enzymes, specific binding molecules, particles, *e.g.* magnetic particles, and the like. Specific binding molecules include pairs, such as biotin and streptavidin, digoxin and antidigoxin etc. For the specific binding members, the complementary member would normally be labeled with a molecule that provides for detection, in accordance with known procedures.

30 A variety of other reagents may be included in the screening assays described herein. Where the assay is a binding assay, these include reagents like salts, neutral proteins, *e.g.* albumin, detergents, etc that are used to facilitate optimal protein-protein binding, protein-DNA binding, and/or reduce non-specific or background interactions. Reagents that improve the efficiency of the assay, such as protease inhibitors, nuclease inhibitors, anti-microbial agents, *etc.* may be used. The components are 35 added in any order that provides for the requisite binding. Incubations are performed at any suitable

temperature, typically between 4 and 40°C. Incubation periods are selected for optimum activity, but may also be optimized to facilitate rapid high-throughput screening. Typically between 0.1 and 1 hours will be sufficient.

Where the screening assay is a binding assay, one or more of the molecules may be joined to a label, where the label can directly or indirectly provide a detectable signal. Various labels include radioisotopes, fluorescers, chemiluminescers, enzymes, specific binding molecules, particles, e.g. magnetic particles, and the like. Specific binding molecules include pairs, such as biotin and streptavidin, digoxin and antidigoxin etc. For the specific binding members, the complementary member would normally be labeled with a molecule that provides for detection, in accordance with known procedures.

The compounds having the desired activity (e.g., reduction of sAC mRNA levels, reduction of sAC polypeptide levels, or reduction of sAC adenylyl cyclase activity) may be administered in a physiologically acceptable carrier to a host for reversible reduction of spermatogenesis and reduction of male fertility. The agents may be administered in a variety of ways, orally, topically, parenterally e.g. subcutaneously, intraperitoneally, by viral infection, intravascularly, intramuscularly, by implantation (see, e.g., U.S. Patent No. 5,733,565), etc. Depending upon the manner of introduction, the compounds may be formulated in a variety of ways. The concentration of active compound in the formulation may vary from about 0.1-100 wt.%.

The pharmaceutical compositions can be prepared in various forms, such as granules, tablets, pills, suppositories, capsules, suspensions, salves, lotions and the like. Pharmaceutical grade organic or inorganic carriers and/or diluents suitable for oral and topical use can be used to make up compositions containing the therapeutically-active compounds. Diluents known to the art include aqueous media, vegetable and animal oils and fats. Stabilizing agents, wetting and emulsifying agents, salts for varying the osmotic pressure or buffers for securing an adequate pH value, and skin penetration enhancers can be used as auxiliary agents. Pharmaceutically acceptable excipients have been amply described in a variety of publications, including, for example, "Remington: The Science and Practice of Pharmacy", 19th Ed. (1995), or latest edition, Mack Publishing Co.

#### ISOLATED HUMAN sAC POLYNUCLEOTIDES

The present invention provides isolated human sAC polynucleotides. These isolated polynucleotides are useful for producing isolated polypeptides of the invention, as described below. They are also useful as hybridization probes for detecting the presence of nucleic acid molecules which hybridize under stringent hybridization conditions to an isolated polynucleotide of the invention. They are further useful in diagnostic methods, including methods to detect a human sAC mRNA in a biological sample, methods to identify polynucleotides having sequence similarity to human sAC

polynucleotides of the invention, methods to detect an alteration in a human sAC polynucleotide sequence in a cell, and methods to identify substances which modulate human sAC mRNA levels in a cell.

In some embodiments, the present invention provides isolated polynucleotides comprising the 5 nucleotide sequence set forth in SEQ ID NO:1, or an identifying sequence thereof. An "identifying sequence" is a contiguous sequence of residues at least about 10 nucleotides (nt) to about 20 nt in length, usually at least about 50 nt to about 100 nt in length, that uniquely identifies the provided sequence. Encompassed in the term "human sAC polynucleotide" are isolated polynucleotides comprising at least about 10, at least about 20, at least about 50, at least about 66, at least about 100, 10 at least about 200, at least about 300, at least about 500, at least about 1000, at least about 1500, at least about 2000, at least about 2500, at least about 3000, at least about 3500, at least about 4000, or at least about 4500 or more contiguous nucleotides of the sequence set forth in SEQ ID NO:1, up to the complete coding sequence. Other fragments of interest are those which encode an antigenic determinant, and which therefore can be used to elicit an immune response in a mammalian host.

15 As used herein, "human sAC polynucleotide" encompasses cDNA and genomic sequences. The term "cDNA" as used herein is intended to include all nucleic acids that share the arrangement of sequence elements found in native mature mRNA species, where sequence elements are exons and 3' and 5' non-coding regions. Normally mRNA species have contiguous exons, with the intervening 20 introns, when present, removed by nuclear RNA splicing, to create a continuous open reading frame encoding the protein. The term "human sAC polynucleotide" further encompasses naturally occurring human polymorphisms.

A genomic sequence of interest comprises the nucleic acid present between the initiation codon and the stop codon, as defined in the listed sequences, including all of the introns that are normally 25 present in a native chromosome. It may further include the 3' and 5' untranslated regions found in the mature mRNA. It may further include specific transcriptional and translational regulatory sequences, such as promoters, enhancers, etc., including about 1 kb, but possibly more, of flanking genomic DNA at either the 5' or 3' end of the transcribed region. The genomic DNA may be isolated as a fragment of 100 kbp or smaller; and substantially free of flanking chromosomal sequence. The genomic DNA flanking the coding region, either 3' or 5', or internal regulatory sequences as sometimes found in 30 introns, contains sequences required for proper tissue and stage specific expression.

The sequence of the 5' flanking region may be utilized for promoter elements, including enhancer binding sites, that provide for developmental regulation in tissues where human sAC is expressed. The tissue specific expression is useful for determining the pattern of expression, and for providing promoters that mimic the native pattern of expression. Naturally occurring polymorphisms 35 in the promoter region are useful for determining natural variations in expression, particularly those

that may be associated with, e.g., abnormal spermatid function, abnormal spermatogenesis, and/or abnormally low numbers of spermatids.

Alternatively, mutations may be introduced into the promoter region to determine the effect of altering expression in experimentally defined systems. Methods for the identification of specific DNA motifs involved in the binding of transcriptional factors are known in the art, e.g. sequence similarity to known binding motifs, gel retardation studies, etc. For examples, see Blackwell et al. (1995) *Mol. Med.* 1:194-205; Mortlock et al. (1996) *Genome Res.* 6:327-33; and Joulin and Richard-Foy (1995) *Eur. J. Biochem.* 232:620-626.

The regulatory sequences may be used to identify cis acting sequences required for transcriptional or translational regulation of expression, especially in different tissues or stages of development, and to identify cis acting sequences and trans-acting factors that regulate or mediate expression. Such transcription or translational control regions may be operably linked to one of the subject genes in order to promote expression of wild type or altered protein, or other proteins of interest in cultured cells, or in embryonic, fetal or adult tissues, and for gene therapy.

Isolated polynucleotides of the invention also include nucleic acids comprising sequences having sequence similarity or sequence identity to the sequence provided in SEQ ID NO:1. A previously identified cDNA sequence of rat sAC is found under GenBank Accession No. AF081941 (SEQ ID NO:3). The human and rat sAC coding regions share 84% nucleotide sequence identity when aligned, as shown in Figure 4. Isolated sAC polynucleotides of the invention share at least about 85%, more preferably at least about 90% or more, sequence identity with the sequence depicted in SEQ ID NO:1. Polynucleotides of the invention also include nucleic acids having sequence similarity or sequence identity to the sequences provided in SEQ ID NO:1. Nucleic acids having sequence similarity are detected by hybridization under low stringency conditions, for example, at 50°C and 10XSSC (0.9 M NaCl/0.09 M sodium citrate) and remain bound when subjected to washing at 55°C in 1XSSC.

Sequence identity can be determined by hybridization under stringent conditions, for example, at 50°C or higher and 0.1XSSC (9 mM NaCl/0.9 mM sodium citrate). Hybridization methods and conditions are well known in the art, see, e.g., U.S. Patent No. 5,707,829. Nucleic acids that are substantially identical to the provided nucleic acid sequences, e.g. allelic variants, genetically altered versions of the gene, etc., bind to the provided nucleic acid sequences (SEQ ID NO:1) under stringent hybridization conditions. By using probes, particularly labeled probes of DNA sequences, one can isolate homologous or related genes. The source of homologous genes can be any species, e.g. primate species, particularly human; rodents, such as rats and mice; canines, felines, bovines, ovines, equines, yeast, nematodes, etc.

"Hybridization" refers to the association of two nucleic acid sequences to one another by hydrogen bonding. Two sequences will be placed in contact with one another under conditions that favor hydrogen bonding. Factors that affect this bonding include: the type and volume of solvent; reaction temperature; time of hybridization; agitation; agents to block the non-specific attachment of the liquid phase sequence to the solid support (Denhardt's reagent or BLOTO); concentration of the sequences; use of compounds to increase the rate of association of sequences (dextran sulfate or polyethylene glycol); and the stringency of the washing conditions following hybridization. See 5 Sambrook, et al., *supra* (1989), Volume 2, chapter 9, pages 9.47 to 9.57.

"Stringency" refers to conditions in a hybridization reaction that favor association of very 10 similar sequences over sequences that differ. For example, the combination of temperature and salt concentration should be chosen that is approximately 120 to 200 °C below the calculated  $T_m$  of the hybrid under study. The temperature and salt conditions can often be determined empirically in preliminary experiments in which samples of genomic DNA immobilized on filters are hybridized to the sequence of interest and then washed under conditions of different stringencies. See 15 Sambrook, et al., *supra*, at page 9.50.

Variables to consider when performing, for example, a Southern blot are (1) the complexity of 20 the DNA being blotted and (2) the homology between the target and the sequences being detected. The total amount of the polynucleotides to be studied can vary a magnitude of 10, from 0.1 to 1  $\mu$ g for a plasmid or phage digest to 10<sup>-9</sup> to 10<sup>-8</sup>  $\mu$ g for a single copy gene in a highly complex eukaryotic genome. For lower complexity polynucleotides, substantially shorter blotting, hybridization, and exposure times, a smaller amount of starting polynucleotides, and lower specific activity of a target 25 polynucleotide can be used. For example, a single-copy yeast gene can be detected with an exposure time of only 1 hour starting with 1  $\mu$ g of yeast DNA, blotting for two hours, and hybridizing for 4-8 hours with a target polynucleotide radiolabeled with 108 cpm/ $\mu$ g. For a single-copy mammalian gene a conservative approach would start with 10  $\mu$ g of DNA, blot overnight, and hybridize overnight in the 30 presence of 10% dextran sulfate using a target polynucleotide radiolabeled with greater than 108 cpm/ $\mu$ g, resulting in an exposure time of ~24 hours.

Several factors can affect the melting temperature ( $T_m$ ) of a DNA-DNA hybrid between the target and sequence of interest, and consequently, the appropriate conditions for hybridization and 30 washing. In many cases the target is not 100% homologous to the fragment. Other commonly encountered variables include the length and total G+C content of the hybridizing sequences and the ionic strength and formamide content of the hybridization buffer. The effects of all of these factors can be approximated by a single equation:

$$T_m = 81 + 16.6(\log 10 C_i) + 0.4[\%G + C] - 0.6(\%formamide) - 600/n - 1.5(\%mismatch),$$

where  $C_i$  is the salt concentration (monovalent ions) and  $n$  is the length of the hybrid in base pairs (slightly modified from Meinkoth and Wahl, (1984) *Anal. Biochem.* 138: 267-284).

In designing a hybridization experiment, some factors affecting nucleic acid hybridization can be conveniently altered. The temperature of the hybridization and washes and the salt concentration during the washes are the simplest to adjust. As the temperature of the hybridization increases (i.e., stringency), it becomes less likely for hybridization to occur between strands that are nonhomologous, and as a result, background decreases. If the labeled probe is not completely homologous with the immobilized fragment (as is frequently the case in gene family and interspecies hybridization experiments), the hybridization temperature must be reduced, and background will increase. The temperature of the washes affects the intensity of the hybridizing band and the degree of background in a similar manner. The stringency of the washes is also increased with decreasing salt concentrations.

In general, convenient hybridization temperatures in the presence of 50% formamide are 42°C for a target polynucleotide with 95% to 100% sequence identity to the sequence to be detected, 37°C for 90% to 95% sequence identity, and 32°C for 85% to 90% sequence identity. For lower percentage sequence identity, formamide content should be lowered and temperature adjusted accordingly, using the equation above. If the homology between the target polynucleotide and the sequence to be detected are not known, the simplest approach is to start with both hybridization and wash conditions which are nonstringent. If nonspecific bands or high background are observed after autoradiography, the filter can be washed at high stringency and reexposed. If the time required for exposure makes this approach impractical, several hybridization and/or washing stringencies should be tested in parallel. Stringent conditions include hybridization in a solution of at least about 5 x SSC at 65°C, or at least about 4 x SSC at 42°C; see, for example, U.S. Patent No. 5,707,829, the disclosure of which is herein incorporated by reference.

Preferably, hybridization is performed using at least 18 contiguous nucleotides of SEQ ID NO:1. That is, when at least 18 contiguous nucleotides of SEQ ID NO:1 is used as a probe, the probe will preferentially hybridize with a nucleic acid or mRNA comprising the complementary sequence, allowing the identification and retrieval of the nucleic acids of the biological material that uniquely hybridize to the selected probe. Probes of more than 18 nucleotides can be used, e.g. probes of from about 25 nucleotides, about 50, about 75 nucleotides, to not more than about 100 nucleotides, but 18 nucleotides generally represents sufficient sequence for unique identification.

sAC mRNA is preferentially expressed in spermatids. The term "preferentially expressed in spermatids" indicates that sAC mRNA is expressed at a level generally at least about 2-fold, usually at least about 10-fold, typically at least about 20-fold or more higher in spermatids than in somatic cells of the body. A convenient reference cell is the Sertoli cell, which does not express sAC mRNA. Whether an sAC mRNA is expressed preferentially in spermatids can be readily determined by those

skilled in the art, by determining the level of sAC mRNA in spermatids, in comparison with sAC mRNA levels in a somatic cell, including, but not limited to, Sertoli cells. Methods of measuring mRNA levels are known in the art, and any of these methods are suitable for use herein.

The nucleic acids of the invention also include naturally occurring variants of the nucleotide sequences, e.g. degenerate variants, allelic variants, etc. Variants of the nucleic acids of the invention are identified by hybridization of putative variants with nucleotide sequences disclosed herein, preferably by hybridization under stringent conditions. For example, by using appropriate wash conditions, variants of the nucleic acids of the invention can be identified where the allelic variant exhibits at most about 25-30% base pair mismatches relative to the selected nucleic acid probe. In general, allelic variants contain 15-25% base pair mismatches, and can contain as little as even 5-15%, or 2-5%, or 1-2% base pair mismatches, as well as a single base-pair mismatch.

The invention also encompasses homologs corresponding to the nucleic acids of SEQ ID NO:1, where the source of homologous genes can be any related species within the same genus or group. Within a group, homologs have substantial sequence similarity, e.g. at least about 80% sequence identity, usually at least 90%, more usually at least 95% between nucleotide sequences. Sequence similarity is calculated based on a reference sequence, which may be a subset of a larger sequence, such as a conserved motif, coding region, flanking region, etc. A reference sequence will usually be at least about 18 contiguous nt long, more usually at least about 30 nt long, and may extend to the complete sequence that is being compared. Algorithms for sequence analysis are known in the art, such as BLAST, described in Altschul et al. (1990) *J. Mol. Biol.* 215:403-10.

The nucleic acid compositions of the subject invention may encode all or a part of the subject polypeptides. SEQ ID NO:2 gives the amino acid translation of the nucleotide sequence given as SEQ ID NO:1. Accordingly, in some embodiments, an isolated polynucleotide of the invention comprises nucleotide sequences which encode a polypeptide comprising at least about 5, at least about 10, at least about 15, at least about 25, at least about 50, at least about 75, at least about 100, at least about 200, at least about 300, at least about 400, at least about 500, at least about 750, at least about 1000, at least about 1250, at least about 1500, or at least about 1600 or more contiguous amino acids of the sequence set forth in SEQ ID NO:2, up to the full length polypeptide. In some of these embodiments, an isolated polynucleotide of the invention comprises nucleotide sequences which encode a human sAC having the amino acid sequence depicted in SEQ ID NO:2. Also encompassed are human sAC polynucleotides encoding variants, fragments and fusion proteins of the aforementioned polypeptides. Accordingly, the invention provides a human sAC polynucleotide which encodes a variants of a human sAC polypeptide, including variants having conservative amino acid substitutions, and fragments thereof; and fusion proteins comprising any one of the aforementioned polypeptides and a heterologous

polypeptide (i.e., a non-human sAC polypeptide, other than the rat sAC polypeptide sequence provided under GenBank accession No. A081941).

Double or single stranded fragments may be obtained of the DNA sequence by chemically synthesizing oligonucleotides in accordance with conventional methods, by restriction enzyme 5 digestion, by polymerase chain reaction (PCR) amplification, etc. For the most part, DNA fragments will be of at least 18 nt, usually at least 25 nt or 50 nt, and may be at least about 100 nt. Such small DNA fragments are useful as primers for PCR, hybridization screening probes, etc. Larger DNA fragments, i.e. greater than 500 nt are useful for production of the encoded polypeptide. For use in amplification reactions, such as PCR, a pair of primers will be used. The exact composition of the 10 primer sequences is not critical to the invention, but for most applications the primers will hybridize to the subject sequence under stringent conditions, as known in the art. It is preferable to choose a pair of primers that will generate an amplification product of at least about 50 nt, preferably at least about 100 nt. Algorithms for the selection of primer sequences are generally known, and are available in 15 commercial software packages. Amplification primers hybridize to complementary strands of DNA, and will prime towards each other.

Also encompassed by the invention are polynucleotides complementary to a human sAC polynucleotide, as defined above. Further encompassed are human sAC antisense polynucleotides and 20 ribozymes. Various derivatives of the antisense sequence may be prepared, where the phosphates may be modified, where oxygens may be substituted with sulfur and nitrogen, the sugars may be modified, and the like. The antisense sequences may be used by themselves or in conjunction with various toxic 25 moieties, such as metal chelates, sensitizers, ribozymes, and the like. Antisense and/or ribozyme sequences may be used to inhibit spermatogenesis. Antisense polynucleotides, and methods of using such, are described in numerous publications, including, e.g., "Antisense Technology: A Practical Approach" Lichtenstein and Nellen, eds. (1997) IRL Press.

Polynucleotides of the invention are isolated and obtained in substantial purity, generally as 30 other than an intact chromosome. Usually, the DNA will be obtained substantially free of other nucleic acid sequences, generally being at least about 50%, usually at least about 90% pure and are typically "recombinant", i.e. flanked by one or more nucleotides with which it is not normally associated on a naturally occurring chromosome.

The DNA sequences are used in a variety of ways. They can be used in methods to detect 35 human sAC mRNA in a biological sample, as described in more detail below.

The human sAC-encoding DNA may also be used to identify expression of the gene in a biological specimen. The manner in which one probes cells for the presence of particular nucleotide sequences, as genomic DNA or RNA, is well established in the literature and does not require 35 elaboration here. mRNA may be isolated from a cell sample, or may be detected without being first

isolated. mRNA may be amplified by RT-PCR, using reverse transcriptase to form a complementary DNA (cDNA) strand, followed by polymerase chain reaction amplification using primers specific for the subject DNA sequences. Alternatively, mRNA sample is separated by gel electrophoresis, transferred to a suitable support, *e.g.* nitrocellulose, nylon, *etc.*, and then probed with a fragment of the 5 subject DNA as a probe. Other techniques, such as oligonucleotide ligation assays, *in situ* hybridizations, and hybridization to DNA probes arrayed on a solid chip may also find use. Detection of mRNA hybridizing to a human sAC sequence is indicative of human sAC gene expression in the sample.

The human sAC nucleic acid sequence may be modified for a number of purposes, 10 particularly where they will be used intracellularly, for example, by being joined to a nucleic acid cleaving agent, *e.g.* a chelated metal ion, such as iron or chromium for cleavage of the gene; or the like.

The sequence of the human sAC locus, including flanking promoter regions and coding 15 regions, may be mutated in various ways known in the art to generate targeted changes in promoter strength, sequence of the encoded protein, *etc.* The DNA sequence or product of such a mutation will be substantially similar to the sequences provided herein, *i.e.* will differ by at least one nucleotide or amino acid, respectively, and may differ by at least two but generally not more than about ten nucleotides or amino acids. The sequence changes may be substitutions, insertions or deletions.

Deletions may further include larger changes, such as deletions of a domain or exon. Other 20 modifications of interest include epitope tagging, *e.g.* with the FLAG system, HA, *etc.* For studies of subcellular localization, fusion proteins with green fluorescent proteins (GFP) may be used. Such mutated genes may be used to study structure-function relationships of human sAC polypeptides with other polypeptides, or to alter properties of the proteins that affect their function or regulation. Such modified human sAC sequences can be used, for example, to generate transgenic animals.

25 Techniques for in vitro mutagenesis of cloned genes are known. Examples of protocols for scanning mutations may be found in Gustin et al., 1993 Biotechniques 14:22 ; Barany, 1985 Gene 37:111-23; Colicelli et al., 1985 Mol Gen Genet 199:537-9; and Prentki et al., 1984 Gene 29:303-13. Methods for site-specific mutagenesis can be found in Sambrook et al., 1989 Molecular Cloning: A 30 Laboratory Manual, CSH Press, pp. 15.3-15.108; Weiner et al., 1993 Gene 126:35-41; Sayers et al., 1992 Biotechniques 13:592-6; Jones and Winistorfer, 1992 Biotechniques 12:528-30; Barton et al., 1990 Nucleic Acids Res. 18:7349-55; Marotti and Tomich, 1989 Gene Anal. Tech. 6:67-70; and Zhu 1989 Anal. Biochem. 177:120-4.

Human sAC polynucleotides can be obtained by chemical or biochemical synthesis, by recombinant DNA techniques, or by isolating the nucleic acids from a biological source. For example, 35 the nucleic acid may be synthesized using solid phase synthesis techniques, as are known in the art.

Oligonucleotide synthesis is also described in Edge et al., *Nature* (1981) 292:756; Duckworth et al., *Nucleic Acids Res.* (1981) 9:1691 and Beaucage & Caruthers, *Tet. Letts* (1981) 22: 1859. Following preparation of the nucleic acid, the nucleic acid is then ligated to other members of the expression system to produce an expression cassette or system comprising a nucleic acid encoding the subject product in operational combination with transcriptional initiation and termination regions, which provide for expression of the nucleic acid into the subject polypeptide products under suitable conditions.

#### ISOLATED HUMAN SOLUBLE ADENYLYL CYCLASE POLYPEPTIDES

The present invention provides isolated human sAC polypeptides. The polypeptides are useful for a variety of purposes, including use in methods of identifying factors that bind to human sAC polypeptides; generating antibodies specific for human sAC polypeptides; and use in methods of identifying substances which modulate human sAC enzyme activity.

The term "human sAC polypeptide" encompasses an amino acid sequence encoded by an open reading frame (ORF) of the human sAC polynucleotides described herein, including the full-length native polypeptide and fragments thereof, particularly biologically active fragments and/or fragments corresponding to functional domains, e.g. a region or domain having adenylyl cyclase activity, etc.; antigenic fragments thereof; and including fusions of the subject polypeptides to other proteins or parts thereof.

As used herein, "human sAC polypeptide" refers to an amino acid sequence of a recombinant or non-recombinant polypeptide having an amino acid sequence of i) a native human sAC polypeptide, ii) a fragment of a human sAC polypeptide, iii) polypeptide analogs of a human sAC polypeptide, iv) variants of a human sAC polypeptide; v) an immunologically active fragment of a human sAC polypeptide; and vi) fusion proteins comprising a human sAC polypeptide. Human sAC polypeptides of the invention can be obtained from a human biological sample, or from any source whether natural, synthetic, semi-synthetic or recombinant. "Human sAC polypeptide" refers to the amino acid sequences of isolated human sAC polypeptide obtained from a human, and is meant to include all naturally-occurring allelic variants, and is not meant to limit the amino acid sequence to the complete, native amino acid sequence associated with the recited protein molecule.

Those skilled in the art will appreciate that changes can be made to the sequences depicted in SEQ ID NO:2 without substantially affecting a function of the human sAC polypeptide. Thus, the term "human sAC polypeptide" encompasses polypeptides with conservative amino acid substitutions compared with the sequences depicted in SEQ ID NO:2. Examples of conservative amino acid substitutions include Ser/Thr; Ala/Val; Leu/Ile; Asp/Glu; and Phe/Tyr. Clearly, other amino acid substitutions, deletions, and insertions can be made to the polypeptide without affecting one or more

functions of the polypeptide. Those skilled in the art, given the guidance provided in the instant specification, can readily determine whether a given function of a human sAC polypeptide is preserved. One such function is adenylyl cyclase activity. Accordingly, in some embodiments of the invention, an isolated human sAC polypeptide has adenylyl cyclase activity.

5 Whether a human sAC polypeptide has adenylyl cyclase activity can be readily determined by those skilled in the art. Any known method to assay adenylyl cyclase activity can be used, as described above. Adenylyl cyclase activity can be measured in cell lysates comprising sAC, using isolated sAC, or in intact cells comprising sAC, as described above.

10 The term "human sAC polypeptide" encompasses a polypeptide comprising from at least about 5, at least about 10, at least about 15, at least about 25, at least about 43, at least about 50, at least about 75, at least about 100, at least about 200, at least about 300, at least about 400, at least about 500, at least about 750, at least about 1000, at least about 1250, at least about 1500, or at least about 1600 or more contiguous amino acids of the sequence set forth in SEQ ID NO:2. In some embodiments, a human sAC polypeptide comprises the entire sequence set forth in SEQ ID NO:2.

15 The human sAC and rat sAC amino acid sequences share about 76.7% amino acid sequence identity, as measured using the BLASTP program 2.0.9 (matrix blosum62; gap open = 9; gap extension = 2; dropoff = 50; wordsize = 3; no filter). Accordingly, also encompassed by the term "human sAC polypeptide" is a polypeptide sharing at least about 77%, more preferably at least about 80 %, even more preferably at least about 85%, even more preferably at least about 90%, even more 20 preferably at least about 95% or more amino acid sequence identity with the sequence depicted in SEQ ID NO:2.

25 A human sAC polypeptide need not be full length in order to retain adenylyl cyclase activity. Accordingly, in some embodiments, a human sAC polypeptide has a C-terminal truncation or internal deletion(s), while retaining adenylyl cyclase activity. Thus, a human sAC polypeptide may comprise, or may consist essentially of, from amino acid 1 to about amino acid 500, from amino acid 1 to about amino acid 490, from amino acid 1 to about amino acid 480, or from amino acid 1 to about amino acid 470, of the sequence provided in SEQ ID NO:2. In one embodiment, a human sAC polypeptide consists essentially of from amino acid 1 to amino acid 469 of the sequence provided in SEQ ID NO:2.

30 Production of isolated human sAC polypeptides

Isolated human sAC polypeptides of the invention can be obtained by any known method, or a combination of such methods, including isolation from natural sources; production by chemical synthesis; and production by standard recombinant techniques.

35 Human sAC polypeptides can be isolated from a biological source, using affinity chromatography, e.g., using antibodies specific for human sAC are immobilized on a solid support.

Alternatively, human sAC polypeptides can be isolated from a human biological source using a combination of anion exchange, and size exclusion chromatography, using a protocol such as that described in Buck et al. (1999) *Proc. Natl. Acad. Sci. USA* 96:79-84.

The polypeptides may be expressed in prokaryotes or eukaryotes in accordance with conventional ways, depending upon the purpose for expression. For large scale production of the protein, a unicellular organism, such as *E. coli*, *B. subtilis*, *S. cerevisiae*, insect cells in combination with baculovirus vectors, or cells of a higher organism such as vertebrates, particularly mammals, e.g. COS 7 cells, CHO cells, HEK293 cells, MA-10 cells, and the like, may be used as the expression host cells. In some situations, it is desirable to express the gene in eukaryotic cells, where the protein will benefit from native folding and post-translational modifications. The polypeptide can then be isolated from cell culture supernatant or from cell lysates using affinity chromatography methods or anion exchange/size exclusion chromatography methods, as described above.

With the availability of the protein or fragments thereof in large amounts, by employing an expression host, the protein may be isolated and purified in accordance with conventional ways. A lysate may be prepared of the expression host and the lysate purified using HPLC, exclusion chromatography, gel electrophoresis, affinity chromatography, or other purification technique.

#### RECOMBINANT VECTORS

The present invention further provides recombinant vectors ("constructs") comprising a polynucleotide of the invention. Recombinant vectors include vectors used for propagation of a polynucleotide of the invention, and expression vectors. The present invention further provides recombinant vectors comprising human sAC polynucleotides of the invention. Recombinant vectors are useful for propagation of the subject human sAC polynucleotides (cloning vectors). They are also useful for effecting expression of a human sAC polynucleotide in a cell. The choice of appropriate vector is well within the skill of the art. Many such vectors are available commercially.

The subject nucleic acid molecules are generally propagated by placing the molecule in a vector. Viral and non-viral vectors are used, including plasmids. The choice of plasmid will depend on the type of cell in which propagation is desired and the purpose of propagation. Certain vectors are useful for amplifying and making large amounts of the desired DNA sequence.

Other vectors are suitable for expression in cells in culture. These vectors will generally include regulatory sequences ("control sequences" or "control regions") which are necessary to effect the expression of a human sAC polynucleotide to which they are operably linked. Still other vectors are suitable for transfer and expression in cells in a whole organism or person.

Expression vectors generally have convenient restriction sites located near the promoter sequence to provide for the insertion of nucleic acid sequences encoding heterologous proteins. A

selectable marker operative in the expression host may be present. Expression vectors may be used for the production of fusion proteins, where the exogenous fusion peptide provides additional functionality, i.e. increased protein synthesis, stability, reactivity with defined antisera, an enzyme marker, e.g.  $\beta$ -galactosidase, etc.

5 Expression cassettes may be prepared comprising a transcription initiation region, the gene or fragment thereof, and a transcriptional termination region. Of particular interest is the use of sequences that allow for the expression of functional epitopes or domains, usually at least about 8 amino acids in length, more usually at least about 15 amino acids in length, at least about 25 amino acids, at least about 45 amino acids, and up to the complete open reading frame of the gene. After 10 introduction of the DNA, the cells containing the construct may be selected by means of a selectable marker, the cells expanded and then used for expression.

The expression cassettes may be introduced into a variety of vectors, *e.g.* plasmid, BAC, YAC, bacteriophage such as lambda, P1, M13, *etc.*, animal or plant viruses, and the like, where the vectors are normally characterized by the ability to provide selection of cells comprising the expression 15 vectors. The vectors may provide for extrachromosomal maintenance, particularly as plasmids or viruses, or for integration into the host chromosome. Where extrachromosomal maintenance is desired, an origin sequence is provided for the replication of the plasmid, which may be low- or high copy-number. A wide variety of markers are available for selection, particularly those which protect against toxins, more particularly against antibiotics. The particular marker that is chosen is selected in 20 accordance with the nature of the host, where in some cases, complementation may be employed with auxotrophic hosts. Introduction of the DNA construct may use any convenient method, *e.g.* conjugation, bacterial transformation, calcium-precipitated DNA, electroporation, fusion, transfection, infection with viral vectors, biolistics, *etc.*

25 HOST CELLS

The present invention further provides host cells, which may be isolated host cells, comprising human sAC polynucleotides of the invention. Suitable host cells include prokaryotes such as *E. coli*, *B. subtilis*, eukaryotes, including insect cells in combination with baculovirus vectors, yeast cells, such as *Saccharomyces cerevisiae*, or cells of a higher organism such as vertebrates, including amphibians 30 (e.g., *Xenopus laevis* oocytes), and mammals, particularly mammals, *e.g.* COS cells, CHO cells, HEK293 cells, MA-10 cells, and the like, may be used as the expression host cells. Host cells can be used for the purposes of propagating a human sAC polynucleotide, for production of a human sAC polypeptide, or in cell-based methods for identifying agents which modulate a level of sAC mRNA and/or protein and/or enzyme activity in a cell.

The subject nucleic acids can be used to generate transgenic animals or site specific gene modifications in cell lines. The modified cells or animals are useful in the study of sAC function and regulation. For example, a series of small deletions or substitutions may be made in the sAC gene to determine the role of different coding regions in spermatogenesis, signal transduction, substrate binding, *etc.*

5 DNA constructs for homologous recombination will comprise at least a portion of the sAC gene with the desired genetic modification, and will include regions of homology to the target locus. Conveniently, markers for positive and negative selection are included. Methods for generating cells having targeted gene modifications through homologous recombination are known in the art. For 10 various techniques for transfecting mammalian cells, see Keown *et al.* (1990) Methods in Enzymology 185:527-537.

For embryonic stem (ES) cells, an ES cell line may be employed, or ES cells may be obtained freshly from a host, *e.g.* mouse, rat, guinea pig, *etc.* Such cells are grown on an appropriate fibroblast-feeder layer or grown in the presence of leukemia inhibiting factor (LIF). When ES cells 15 have been transformed, they may be used to produce transgenic animals. After transformation, the cells are plated onto a feeder layer in an appropriate medium. Cells containing the construct may be detected by employing a selective medium. After sufficient time for colonies to grow, they are picked and analyzed for the occurrence of homologous recombination. Those colonies that show homologous recombination may then be used for embryo manipulation and blastocyst injection. Blastocysts are 20 obtained from 4 to 6 week old superovulated females. The ES cells are trypsinized, and the modified cells are injected into the blastocoel of the blastocyst. After injection, the blastocysts are returned to each uterine horn of pseudopregnant females. Females are then allowed to go to term and the resulting litters screened for mutant cells having the construct. By providing for a different phenotype of the blastocyst and the ES cells, chimeric progeny can be readily detected. The chimeric animals are 25 screened for the presence of the sAC gene and males and females having the modification are mated to produce homozygous progeny. The transgenic animals may be any non-human mammal, such as laboratory animals, domestic animals, *etc.* The transgenic animals may be used to determine the effect of a candidate drug on spermatogenesis in an *in vivo* environment.

30 ANTIBODIES

The present invention provides antibodies, which may be isolated antibodies, specific for human sAC polypeptides of the invention. Such antibodies are useful, for example, in methods of detecting the presence of a human sAC polypeptide in a biological sample, and in methods of isolating a human sAC polypeptide from a biological sample.

Isolated human sAC polypeptides of the invention are useful for the production of antibodies, where short fragments provide for antibodies specific for the particular polypeptide, and larger fragments or the entire protein allow for the production of antibodies over the surface of the polypeptide. Accordingly, the invention provides isolated antibodies which specifically bind a human sAC polypeptide, or antigenic fragment thereof. Antibodies may be raised to the wild-type or variant forms. Antibodies may be raised to isolated peptides corresponding to these domains, or to the native protein. Antibodies may be raised to polypeptides and/or peptide fragments of sAC from other species. These antibodies may cross-react with human sAC polypeptides. For example, antibodies can be raised using as an immunogen a rat sAC peptide fragment, e.g., amino acids 1518-1607, amino acids 569-593, and/or amino acids 92-108 of the rat sAC amino acid sequence depicted in Figure 2.

The human sAC polypeptides of the invention are useful for the production of antibodies, where short fragments provide for antibodies specific for the particular polypeptide, and larger fragments or the entire protein allow for the production of antibodies over the surface of the polypeptide. As used herein, the term "antibodies" includes antibodies of any isotype, fragments of antibodies which retain specific binding to antigen, including, but not limited to, Fab, Fv, scFv, and Fd fragments, fusion proteins comprising such antibody fragments, and chimeric antibodies. "Antibody specificity", in the context of antibody-antigen interactions, is a term well understood in the art, and indicates that a given antibody binds to a given antigen, wherein the binding can be inhibited by that antigen or an epitope thereof which is recognized by the antibody, and does not substantially bind to unrelated antigens. Methods of determining specific antibody binding are well known to those skilled in the art, and can be used to determine the specificity of antibodies of the invention for a human sAC polypeptide.

Antibodies are prepared in accordance with conventional ways, where the expressed polypeptide or protein is used as an immunogen, by itself or conjugated to known immunogenic carriers, e.g. KLH, pre-S HBsAg, other viral or eukaryotic proteins, or the like. Various adjuvants may be employed, with a series of injections, as appropriate. For monoclonal antibodies, after one or more booster injections, the spleen is isolated, the lymphocytes immortalized by cell fusion, and then screened for high affinity antibody binding. The immortalized cells, i.e. hybridomas, producing the desired antibodies may then be expanded. For further description, see *Monoclonal Antibodies: A Laboratory Manual*, Harlow and Lane eds., Cold Spring Harbor Laboratories, Cold Spring Harbor, New York, 1988. If desired, the mRNA encoding the heavy and light chains may be isolated and mutagenized by cloning in *E. coli*, and the heavy and light chains mixed to further enhance the affinity of the antibody. Alternatives to *in vivo* immunization as a method of raising antibodies include binding to phage display libraries, usually in conjunction with *in vitro* affinity maturation.

## COMPOSITIONS

The present invention further provides compositions, including pharmaceutical compositions, comprising the polypeptides, polynucleotides, recombinant vectors, host cells, and antibodies of the invention. These compositions may include a buffer, which is selected according to the desired use of the polypeptide, polynucleotide, recombinant vector, host cell, or antibody, and may also include other substances appropriate to the intended use. Those skilled in the art can readily select an appropriate buffer, a wide variety of which are known in the art, suitable for an intended use. In some instances, the composition can comprise a pharmaceutically acceptable excipient, a variety of which are known in the art and need not be discussed in detail herein. Pharmaceutically acceptable excipients have been amply described in a variety of publications, including, for example, "Remington: The Science and Practice of Pharmacy", Alfonso R. Gennaro (latest edition), Lippincott, Williams, and Wilkins.

## EXAMPLES

The following examples are put forth so as to provide those of ordinary skill in the art with a complete disclosure and description of how to make and use the present invention, and are not intended to limit the scope of what the inventors regard as their invention nor are they intended to represent that the experiments below are all or the only experiments performed. Efforts have been made to ensure accuracy with respect to numbers used (e.g. amounts, temperature, etc.) but some experimental errors and deviations should be accounted for. Unless indicated otherwise, parts are parts by weight, molecular weight is weight average molecular weight, temperature is in degrees Celcius, and pressure is at or near atmospheric.

## EXAMPLE 1

Genomic structure of the human soluble adenylyl cyclase gene

A human testis library was screened with a rat cDNA sequence corresponding to the catalytic and regulatory domains of sAC. Five positive clones were isolated. Two PAC clones from human chromosome 1q24 were sequenced in their entirety. The human sAC gene comprises 32 exons distributed over 120 kb of genomic DNA. The 4896-nucleotide sequence encoding human sAC is shown in Figure 3, and is set forth in SEQ ID NO:1. An alignment of human and rat (GenBank Accession No. AF081941; SEQ ID NO:3) sAC-encoding nucleotide sequence, provided in Figure 4, shows that these two sequences share 84% nucleotide sequence identity.

The human sAC amino acid sequence is depicted in Figure 1. The C1 domain is encoded by exons 2-6, and the C2 domain is encoded by exons 8-11. The encoded human sAC polypeptide has 1614 amino acids (SEQ ID NO:2). An alignment of the 1608-amino acid rat and 1614-amino acid human amino sequences is shown in Figure 2. The human sAC and rat sAC amino acid sequences

were aligned using the BLASTP program 2.0.9 (matrix file: blosum50; gap open = 9; gap extension = 2; dropoff = 50; wordsize = 3; no filter). Using these parameters, it was determined that the human sAC amino acid sequence shares 76.7% amino acid sequence identity with the rat sAC amino acid sequence (SEQ ID NO:4), as shown in Figure 2. The C1 and the C2 domains, together with exon 13, 5 which encodes the P loop, are the most highly conserved regions between rat and human sAC. The C-terminal half of human sAC protein is less conserved, with the exception of the leucine zipper domain, and a region homologous to a tetratricopeptide repeat, a 34-amino acid domain involved in protein-protein interactions.

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## EXAMPLE 2

### Bicarbonate ions activate sAC adenylyl cyclase activity

Experiments were conducted to determine whether bicarbonate ions affect sAC adenylyl cyclase activity. The results are shown in Figure 5. HEK-293 cells were transfected with either 20  $\mu$ g of empty plasmid (“mock transfected cells”), C1C2-recombinant human sAC (“C1-C2 Rec. Hum sAC”), or full-length human sAC (FL Rec. Hum sAC”) constructs. After harvesting and homogenization of the cells, the 100,000  $\times$  g supernatant was used for the assay. Adenylate cyclase activity was measured in the presence (solid bars) or absence (stippled bars) of 50 mM bicarbonate. Supernatants extracted from human sperm were used as a source of native human sAC (“Human Sperm AC”). Data are expressed as picomoles of cAMP formed per minute per milligram of total 20 cellular protein. The data shown in Figure 5 demonstrate that human sAC adenylyl cyclase activity is activated by bicarbonate.

While the present invention has been described with reference to the specific embodiments thereof, it should be understood by those skilled in the art that various changes may be made and equivalents may be substituted without departing from the true spirit and scope of the invention. In addition, many modifications may be made to adapt a particular situation, material, composition of matter, process, process step or steps, to the objective, spirit and scope of the present invention. All such modifications are intended to be within the scope of the claims appended hereto.

## CLAIMS

What is claimed is:

1. An isolated polypeptide comprising at least about 43 contiguous amino acids of the 5 sequence depicted in SEQ ID NO:2.
2. The isolated polypeptide of claim 1, wherein said polypeptide has an amino acid sequence at least about 78% identical to the amino acid sequence depicted in SEQ ID NO:2.
- 10 3. The isolated polypeptide of claim 1, wherein said polypeptide has the amino acid sequence depicted in SEQ ID NO:2
4. The isolated polypeptide of claim 1, wherein said polypeptide has manganese-dependent soluble adenylyl cyclase activity.
- 15 5. A composition comprising the polypeptide of claim 1, and a buffer.
6. An isolated polynucleotide which encodes a polypeptide comprising at least about 6 contiguous amino acids of the sequence depicted in SEQ ID NO:2.
- 20 7. The isolated polynucleotide of claim 6, wherein said polynucleotide encodes a polypeptide having the sequence depicted in SEQ ID NO:2.
8. The isolated polynucleotide of claim 6, wherein said polynucleotide has a nucleotide 25 sequence at least about 85% identical to the sequence depicted in SEQ ID NO:1.
9. The isolated polynucleotide of claim 8, wherein said polynucleotide hybridizes, under stringent conditions, with the polynucleotide sequence depicted in SEQ ID NO:1.
- 30 10. The isolated polynucleotide of claim 9, wherein said polynucleotide has the nucleotide sequence depicted in SEQ ID NO:1.
11. An isolated polynucleotide comprising at least about 66 contiguous nucleotides of the sequence set forth in SEQ ID NO:1.

12. The isolated polynucleotide of claim 11, wherein the polynucleotide encodes a manganese-dependent soluble human adenylyl cyclase.
13. A recombinant vector comprising the polynucleotide of claim 6.
- 5
14. An isolated host cell comprising the polynucleotide of claim 13.
15. The isolated host cell of claim 14, wherein the cell is a prokaryotic cell.
- 10
16. The isolated host cell of claim 14, wherein the cell is a eukaryotic cell.
17. An antibody which specifically binds a polypeptide comprising at least about 6 contiguous amino acids of the sequence set forth in SEQ ID NO:2.
- 15
18. The antibody of claim 17, wherein said antibody is attached to a detectable label.
19. The antibody of claim 17, wherein said antibody is attached to a solid support.
20. A method of identifying a substance which modulates human soluble adenylyl cyclase activity, comprising the steps of:
  - (a) contacting human soluble adenylyl cyclase with a substance to be tested, forming a test sample;
  - (b) measuring adenylyl cyclase activity in the test sample, wherein an effect on adenylyl cyclase activity in the test sample compared to a control sample in the absence of the substance being tested is an indication that the substance modulates adenylyl cyclase activity.
- 25
21. The method of claim 20, wherein said adenylyl cyclase is present in a cell lysate.
22. The method of claim 20, wherein said adenylyl cyclase is present in an intact cell, and wherein said adenylyl cyclase is encoded by nucleotide sequences comprised within an exogenous expression vector.
- 30
23. A method of inhibiting spermatogenesis in a male, comprising administering to the male a substance which preferentially inhibits soluble adenylyl cyclase activity.

24. The method of claim 23, wherein the male is a human.
25. The method of claim 24, wherein the male is a rodent.
- 5 26. A reversible method of male contraception, comprising administering to a male a pharmaceutical composition comprising an effective amount of a substance which preferentially inhibits soluble adenylyl cyclase.
- 10 27. The method of claim 26, wherein the male is a human male.
28. The method of claim 26, wherein the male is a rodent.
- 15 29. A method of reducing the number of motile sperm produced by a male, comprising administering to a male a pharmaceutical composition comprising an effective amount of a substance which preferentially inhibits soluble adenylyl cyclase.

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**FIG. 1**

Human sAC deduced amino acid sequence

MNTPKEEFQDWPIVRIAAHLPDLIVYGHFSUPERFMDYFDGVLMFVDISGFTAMTEKFSSAMYMDRGAE  
QLVEILNYHISAIKEVLIFFGGDILKFAGDALLALWRVERKQLKNIITVVIKCSLEIHGLFETQEWEEG  
LDIRVKIGLAAGHISMLVFGDETHSHFLVIGQAVDDVRLAQNMADMVLSPNCWQLCDRSMIEIESV  
PDQRAVKVNFLKPPPNFNDEFFTCKCTFMHYYPSEHKNLLRLACTLKPDPPELEMSLQKYVMESILKQ  
IDNKQLQGYLSELRPVTIVFVNLMFEDQDKAEEIGPAIQDAYMHITSVLKIFQGQINKVFMFDKGCSFL  
CVFGFPGEKVPDELTHALECAMDIFDFCSQVHKIQTVSIGVASGIVFCGIVGHTVRHEYTVIGQKVNL  
ARMMMYYPGIVTCDSVTYNGSNLPAYFFKELPKKVMKGVADSGPLYQYWRTEKVMFGMACLICNRKED  
YPLLGRNKEINYFMYTMKKFLISNSSQVLMYEGLPGYGKSQILMKIEYLAQGKNHRIIAISLNKISFHQ  
TFYTIQMFMANVLGLDTCKHYKERQTNLRNKVMTLLDEKFYCLLNDIFHVQFPIREISRMSTLKKQKQ  
LEILFMKILKLIVKEERIIFIIDEAQFVDSTSWRFMEKLIRTLPIFIIMSLCPFVNIPCAAARAVIKNR  
NTTYIVVGAQPNDISNKICLDLNVSCISKELDSYLGEGSCGIPFYCEELLKNLEHHEVLVFQQTESEE  
KTNRTWNNLFKYSIKLTEKLNMTLHSDKESEEVCHLTSGVRLKNLSPPTSLKEISLIQLDSMRLSHQM  
LVRCAAIIGLTFTTELLFEILPCWNMKMMIKTLATLVESNIFYCFRNGKELQKALKQNDPSFEVHYRSL  
SLKPSEGMDHGEQQQLRELENEVIECHRIRFCNPMMQKTAYELWLKDQRKAMHLKCARFLEEDAHRCDH  
CRGRDFIPYHHFTVNIRLNALDMAIKKMAMSHGFKTEEKLILSNSEIPETSAFFPENRSPEEIREKIL  
NFFDHVLTMKTSDEDIIPLESCQCCEEILEIVILPLAHHFLALGENDKALYYFLEIASAYLIFCDNYMA  
YMYLNEGQKLLKTLKKDKWSQTFSATFYSLKGEVCFNMGQIVLAKKMLRKALKLNRIFPYNLISLF  
LHIHVEKNRHFHYVNRQAQESPPPGKKRLAQLYRQTVCLSLLWRIYSYLFHCKYYAHLAVMMQMNTA  
LETQNCFQIIKAYLDYSLYHHLAGYKGWFKYEVMAEHIFNLPLKGEGIEIVAYVAETLVFNKLIMGH  
LDLAIELGSRALQM WALLQNPNRHYQSLCRLSRC LLSRYPQLIQVLGRLWELSVTQEHI FSKAFFYF  
VCLDILL YSGFVYRTFEECLEFIHQYENNRLKFHGLLGLYSSVAIWYARLQEWDNFYKFSNRAKNL  
LPRRTMTLTYDGISRYMEGQLHLQKQIKEQSENAQASGEELLKNLENLVAQNTTGPVFCPRLYHLMA  
YVCILMGDGQKCGFLNTALRLSETQGNILEKCWLNMNKE SWYSTSELKEDQWLQTI LSLPSWEKIVAG  
RVNIQDLQKNKFLMRANTVDNHF

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**FIG. 2A**

Alignment of human and rat sAC amino acid sequences

rsAC	1	MSARRQELQDRAIVKIAAHL PDLIVYGD[F]SPERPSV[K]FDGVL MFVDISGFTAMTEKFSTAMYMDRGAEQ
hsAC	1	MNTPKEEFQDWPIVRIA AHL PDLIVYGH[F]SPERPFMDYFDGVL MFVDISGFTAMTEKFSSAMYMDRGAEQ
rsAC	71	LVEILNYYISAI VEKVLIFGGDILK FAGDALL ALW[K]VERKQLKNIITVVIKCSLEIHGLFEAK[E]EEGLD
hsAC	71	LVEILNYHISAI VEKVLIFGGDILK FAGDALL ALW[K]VERKQLKNIITVVIKCSLEIHGLFETQE[E]EEGLD
rsAC	141	IRVKIGLAAGHITMLVFGDETRNYFLVIGQAVDDVRLAQNMNDVILSPNCWQLCDRSMIEIERIPDQ
hsAC	141	IRVKIGLAAGHISMLVFGDETHSHFLVIGQAVDDVRLAQNMNDVILSPNCWQLCDRSMIEIESVPDQ
rsAC	211	RAVKV[FLKPPP]TFNFDEFFAKCMAFMDYYPSGDHKN[FLRLACM]ESDPELE[SLQKYVMEI]ILKQIDDK
hsAC	211	RAVKV[FLKPPP]TFNFDEFFTKCTTFMHHYPSGEHKNLLRLACTL[KPDPELEM]SLQKYVMEISILKQIDNK
rsAC	281	QLRGYLSELRPVTIVFVNLMFKEQDKAEVIGSAIQAACVHITSVLKVFRGQINKVFMFDKGCSFLCVFGF
hsAC	281	QLQGYLSELRPVTIVFVNLMFEDQDKAEIIGPAIQDAYMHTSVLKI[FG]QINKVFMFDKGCSFLCVFGF
rsAC	351	PGEKAPDEI[THALES]AVDIFDFCSQVHKIRTVSIGVASGIVFCGIVGHTVRHEYTVIGQKVNI[AARMMMY]
hsAC	351	PGEKVPDEL[THALEC]AMDIFDFCSQVHKI[Q]TVSIGVASGIVFCGIVGHTVRHEYTVIGQKVNI[AARMMMY]
rsAC	421	YPGIVTCDSVTYDGSNLPAYFFKELPKVMKGVADPGPVYQCLGLNEKVMFGMAYLICNRYEGYPPLLGRV
hsAC	421	YPGIVTCDSVTYNGSNLPAYFFKELPKVMKGVADSGPLYQYWGRTEKVMFGMACLICNRKEDYPPLLGRN
rsAC	491	REIDYFMSTMKDFLMTNC[SRVLMYEGLPGYGSQVLM]EIEYLA[SQHENHRA]MAIALTKISFHQNFTI[QI]
hsAC	491	KEINYFMYTMKKFLISNSSQVLMYEGLPGYGSQVLMKIEYLAQGK[NHRIIAISLNKISFHQTFTI]QM
rsAC	561	L[MANVLGLDTCKHYKERQTNL]QNRMK[TL]DDK[YHCLLN]DIFHVQFPV[SR]MSKIRKQKQLEALFMKI
hsAC	560	FMANVLGLDTCKHYKERQTNLRNKV[MTLLDEKFYCLLN]DIFHVQFP[ISREI]SRMSTLKKQKQLEILFMKI
rsAC	631	LEQTMRERIIIFIIDEAQFVDVA[SWAFIEKL]IRSMP[IFIV]MSLCPFPETPCAAANAI[MKNRNTTYITL]GT
hsAC	630	LKLI[MRERIIIFIIDEAQFVDST]SWRFMEKLIRTLP[IFI]IMSLCPFVNIPCAAARAVI[KNRNTTYIVMGA]
rsAC	701	MQPQEIRD[KV]CVDLSVSSIPRELD[SYL]VEGSCGIPYYCEELLKNLDHHR[IL]IFQQAEAEKTNVITWNNLF
hsAC	700	VQPNDISNKICL[DLNN]VSCISKE[LD]SYLGE[GSCGIPFYCEELLKNL]FQQTESEEKTNRTWNNLF
rsAC	771	KYSVKPTEDM[YL]YT[TSIAAGQ-KEACY]LTSGVRLKNLSPPIASLKEISL[Q]LDSMSL[SHQMLVRCAAII]GLT
hsAC	770	KYSIKLTEKL[NM]VTLHSDKE[SEEVCH]LTSGVRLKNLSPPT[SLKEISLI]QLD[SMR]L[SHQMLVRCAAII]GLT
rsAC	840	FTTELLFEILPCWNMKMMIK[AL]ATL[VESNM]DCFRSSKDLQLALKQNVTTFEVHYRSLSLKSK[E]LAYSE
hsAC	840	FTTELLFEILPCWNMKMMIK[TL]ATL[VESN]IFYC[FRNG]KELQKALKQN[DP]SFEVHYRSLSLK[PGMDH]GE
rsAC	910	EEQLREMEGEVIECRI[LR]FCRPI[MQ]K[TAYELWLKDQ]KKV[HLK]CARFLEE[SAHRCN]HCRN[RDF]IPYHHFT
hsAC	910	EEQLRELENEVIECHR[IR]FCN[PM]MQK[TAYELWLKDQ]RKAMHLKCARFLEE[DAHRC]DHCRG[RDF]IPYHHFT
rsAC	980	ADIRLN[TL]DMDT[VK]KMKV[SHGFKTE]DEVIFSK[SEI]PR-KFKFPEN[ISI]TETREKILHFFDNVII[IKMRTSQ]
hsAC	980	VNIRLN[AL]DMAIKK[MA]SHGFKTE[KL]L[NS]EIP[TS]AFFPEN[RS]P[EEI]REKILNFFDHVLTKMKTSD
sAC	1049	DDVIPLESCH[CE]ELLQIVILPLAQHFVALEEN[KALYYF]LEASAYL[IL]GDNYNA[MYL]GEGERLLKSLT
hsAC	1050	EDIIPLESQ[CEE]ILEIIVILPLAHHFLALGEND[KALYYF]LEIASAYL[IF]CDNYNA[MYL]NEGQKLLKTLK
sAC	1119	NEDSW[WSQT]FEYATFYSLKGE[CFNM]GQMVLAKKMLR[KALKL]NRMFPCNL[SL]TFQ[HM]IEKNRLSHFMNQ
hsAC	1120	KDKSWSQT[FE]SATFYSLKGE[CFNM]GQIVLAKKMLR[KALKL]NRI[FPY]NL[ISL]FLHIHVEKNRHFHYVNR

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Alignment of human and rat sAC amino acid sequences

**FIG. 2B**

rsAC 1189	HTQEGLPGKK	LAQLFLQSSCF	SLLW	KIYSLN	FFFFHY	KYYGRLAA	IMQMNTS	LETQNN	FQIIKAF	LDFS							
hsAC 1190	QAQE	SPPPGKKR	LAQLYRQ	QTIVC	SLLW	RISYSY	LFHCKYYA	HLAMM	QMNTA	LETQNC	FQIIKAY	DMS					
rsAC 1258	LYRHLAGY	EVWFKYE	ILV	MEQ	LN	NPLKGE	AFE	IMAYA	ADALGH	HIKF	TGH	LDLAI	ELGSRA	HKMWSLL			
hsAC 1260	LYHHLAGY	KGVWFKYE	VMAMEH	IF	NL	PLKGE	GEGIE	IMAYA	YMAETL	VFNKL	IMG	HLDLAI	ELGSRA	LQMWALL			
rsAC 1328	RNPNKYHMV	L	CRLSKP	LFL	KSRY	KHL	VQLGWL	WDL	SVT	EEH	IFSKAFFY	FVCLD	IMLYSGF	IYRTFEEC			
hsAC 1330	QNPNRHYQS	L	CRLSRC	L	NSRYPQ	L	IQVLGRL	WEI	SVT	QE	IFSKAFFY	FVCLD	IMLYSGF	VYRTFEEC			
rsAC 1398	LEFI	IHNED	NRILKF	QSGLLL	GLY	SCI	AMWYARLQ	EWDNFY	KFSNRAK	TL	MTRRT	PTV	YYEG	ISRYMEG			
hsAC 1400	LEFI	HQYEN	NRILKF	HSGLLL	GLY	SSM	AIWYARLQ	EWDNFY	KFSNRAK	N	LPRT	MTL	TYYDG	ISRYMEG			
rsAC 1468	QVLHLQKQI	EEQ	QAENA	QDSG	VELL	KALE	TLV	AQNTTGPV	FY	PRL	YHLMAY	V	CILMGDG	HSCDFFLNTA			
hsAC 1470	QVLHLQKQI	KEQ	SENA	QASGE	ELL	KNL	ENL	V	AQNTTGPV	F	PRL	YHLMAY	V	CILMGDG	QKCGLFLNTA		
rsAC 1538	LSETQGN	LEKCWL	SMSKE	WWYS	SAPEL	T	GDQWL	QT	V	SLPSW	DKIVSGN	VT	QDVQ	QKNKFL	MRVN	I	LDNP
hsAC 1540	LSETQGN	LEKCWL	N	MKESW	YSTSEL	K	EDQWL	QT	I	SLPSW	EKIVAG	RVNI	QDL	QKNKFL	MR	ANT	VDNH
rsAC 1608	F																
hsAC 1610	F																

Human sAC cDNA sequence

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**FIG. 3A**

ATGAACACTCCAAAAGAAGAATTCCAGGACTGGCCCAGTCAGAATAGCAGCTCATTACAGACCTCATTGTCTATGGACA  
 TTTCTCCCCAGAGCGACCCTTATGGATTATTTGACGGAGTCCTGATGTTGTTGATATTCAAGGTTTACTGCAATGACTG  
 AGAAGTTCAGCAGTGCATGTACATGGACAGAGGGCTGAGCAGTTGGTGGAGATCCTCAACTACCACATAAGTCAATAGT  
 GAGAAAGTGTGATTTGGAGGAGACATCCTGAAATTGCAAGGTGATGCACTGCTAGCCCTGTGGAGGGTGGAGCGAAAGCA  
 GCTGAAAAACATTATCACAGTGTAAATTAAATGTAGCCTGGAGATCCATGGATTGTTGAGACCCAGGAGTGGGAAGAAGGCC  
 TAGACATCCGAGTCAAGATAGGACTGGCTGCTGCCACATCAGCATGTTGGCTTGAGATGAAACACACAGCCACTTCTG  
 GTGATTGGTCAGGAGTGGACGATGCGCCTGCCAGAACATGGCTCAGATGAATGATGTTATTCTGTCACCAAACACTGCTG  
 GCAGCTCTGTGACCGGAGCATGATTGAAATTGAGAGTGTCCAGATCAGAGAGCAGTTAAGGTTAACTTCTTAAACACCACCC  
 CCAATTAAATTTGATGAATTTCACAAAGTGTACGACCTCATGCATTATTATCCTCTGGTGGACACAAAAACCTCCTG  
 AGGCTTGCATGCACGCTGAAGCCTGATCCTGAACTGGAGATGTCCTACAAAAGTATGTGATGAAAGCATTGAAAGCAGAT  
 TGATAACAAACAGCTTCAGGGCTATTATCTGAGCTTCGCCCAGTGACGATTGTTGAAACCTGATGTTGAAGACCAAG  
 ACAAAAGCAGAAGAGATAGGCCAGCCATCAGGATGCCATATGCACATCACTCTGTCCTGAAAGATCTTCCAAGGCCAAATC  
 AATAAAGTCTCATGTTGACAAGGCTGCTCTTCCTCTGTGTCCTGGCTCCCTGGGAAAAGGTACCTGACGAGCTCAC  
 TCATGCTCTGGAATGTCTGATGGATATATTGACTTCTGCTCTCAAGTCCACAAACTGTATCCATCGGTGTTGCCA  
 GTGGGATTGTTCTGTTGAGACACTGTGAGACACAGTACACAGTCATTGGTCAAAAGTCAACTTAGCTGCC  
 AGGATGATGATGTTACTACCCAGGAATTGAGCTGACCTGCTCACCTACAATGGGAGCAACCTACCAGCTACTTTTAA  
 AGAGCTTCAAAGAAAGTTATGAAAGGTGTTGAGATTCTGGACCATTGTATCAGTATTGGGGCGTACTGAGAAAGTCATGT  
 TTGGTATGGCGTGCCTCATCTGCAACAGAAAGGAGGATTACCCCTTGCTGGACGTAATAAAGAGATCAACTACTCATGTAT  
 ACTATGAAAGAAATTGATATCTAACAGCAGCCAAGTCTTAATGTATGAGGGATTACCAAGGATATGGAAAAGCCAGATACT  
 TATGAAAATTGAGTACCTGCCAAGGTAAAGAATCACAGGATTATTGCCATTGAAATAAGATCAGCTTCCATCAAACCT  
 TCTATACCATCCAGATGTTCATGCCAATGTCCTAGCCCTAGACACTTGTAAACATTATAAAGAACAGACAGAACCTCGA  
 AATAAAGTCATGACACTGTTGGATGAAAAGTCTACTGTCTCTTAATGACATTTCATGTCAGTCCCTATTCTCGGGA  
 GATTTCAGGATGAGCACCTGAAAAAGCAAAACATTGAAATATTGTTATGAAAGATCTGAGCTGATAGTGAAGAG  
 AAAGGATTATTGATCATTGAGGGCCAGTTGTTGAGCTGACCTCCTGGAGATTATGGAGAAGCTTATCCGGACTCTT  
 CCTATCTCATCATTATGTCCTGTCCTCGTTAACATTCCCTGTGAGCTGCCAGGGCGTAATAAAGAACAGGAACAC  
 CACCTACATTGTCGTTGGCAGTACAGCCTAACGACATCTCCAACAAGATCTGCTTGCACCTCAATGTGAGCTGCATCTCA  
 AAGAACTGGACTCGTACCTGGGGAGGGAGCTGTTGAGCTGAGGAAATAGGACCTGGAATAACCTGTTCAAGTATTCCATTAAAGCT  
 AACAGAGAAGTTAACATGGTTACTCTCATAGTGATAAGGAAAGTGAAGAAAGTCTGTCACCTCACAGTGGCGTCAAGACTGA  
 AAAACCTGTCACCTCAAACGTCAATTAAAGAAATCTCTGATCCAGCTGGATAGCATGAGACTTTCCCACCAAATGCTGGT  
 AGATGTGCTGCCATCATTGGCCTGACCTCACCACTGAGTTGTTGAGATTCTCCCTGTTGGAATATGAAAGATGATGATGAT  
 CAAGACCCCTGGCAACCCCTAGTGGAACTAACATTTTTATTGTTCCGGAAATGGCAAGGAGCTTCAAAAGGCCCTGAACAGA  
 ATGATCCCTCATTTGAGGTGCACTATGTTCTGTCCTGTCAGCTGAAAGCCAGTGAAGGGATGGATCACGGTGAAGAGGAACAGCTT  
 CGTGAACCTGGAGAAATGAGGTGATCGAGTGGCACAGGATTGATTGTAACCCTATGATGAGAAAACAGCCTACGAGCTGTG  
 GCTCAAGGACCAGAGAAAAGCCATGCACTGAAATGTGCCGCTTTAGAAGAAGATGCCACAGATGTGACCACTGCCGAG  
 GCAGGGACTTCATTCCCTATCATCCTCACAGTGAATATTGGCTCAACGCTTACGATGGATGCCATTAAAAGATGGCT  
 ATGTCATGGATTAAAAGCTGAAGAAAAGCTTATCTGAAATTCTGACCTGAGGTTACCAACTCAGAGATTCCCTGAGACATCTG  
 TGCATTGTCATTGAAAGACATCTGACGAAGACAATCTGACGAAGACA  
 TTATCCCTCTGGAATCTGCCAGTGTGAAGAAATCCTAGAGATTGTCATCTGCTCTGGCCACCATTTCTGGCTTTGGGA  
 GAAAATGACAAGCCTTATATTACTCTTAGAAATTGATCTGCTTATCTCATCTTGTGATAACTACATGCCATACATGTA  
 TTTGAATGAAGGACAGAAGTTGCTAAAAGCTCAAGAAGGACAAATCTGAGCTGCTTACGAGGAGACATTGAGTCTGCCACCTTACA  
 GCCTCAAAGGTGAGGTCTGTTCAATATGGCCAGATAGTGCCTGCAAGAAAATGCTGAGGAAGGACTGAAGCTCCTCAAC  
 CGAATCTTCCCTACAACCTAACATTCTGTTCTCCATATCCATGTCGAGAAAAACAGACACTTCAATTGTAATCGGCA  
 GCCCAAGAGAGCCCACCTCCAGGAAGAAGAGGCTGGCACAACATTACCGCAAACGCTGCTGCCCTTGTGGCGCA  
 TCTATAGCTACAGTTATCTTCACTGCAAGTATTATGCCACCTGGAGTTATGATGCAAATGAATACTGCACTGGAAACT  
 CAAAATTGTTCCAGATCATTAAGGCTTACCTAGACTATTGCTATACCACCTGGCTGGCTACAAAGGTGTGGTTCAA  
 ATATGAAGTCATGCCATGGAGCACATCTCAACCTCCCCCTGAAAGGGAGGGATTGAAATGTCGGCATACGTGGCTGAGA  
 CACTGGCTTCAACAAGCTCATATGGACACCTGGATTGGCATTGAGTTAGGCTCCCGAGCCCTCAGATGTGGGCAC  
 CTCCAGAACCCACCGACATTATGTCAGACTTAGCAGATGTCCTCTGAAACAGCAGATAACCGCAATTGAT  
 CCAGGTGCTGGGGCGCTGTGGAGCTTCTGTAACACAGGAACACATCTCAGCAAGGATTGTTCTATTGTCGTTGG  
 ACATCCTGCTTATTCTGTTTGTGTTATAGAACATTGAGAATGTTGAAATTGATACACCAATACGAAACACA  
 CTCAGTCCACAGTGGACTCCTCTGGACTTATTCTCTGAGCTATCTGGTATGCCAGACTCAGGAATGGGACA  
 TTACAAATTTCACAGTGGACTCCTCTGGACTTATTCTCTGAGCTATCTGGTATGCCAGACTCAGGAATGGGACA  
 TGAGGGCAAGTTCTCACCTCAAAACAAATCAAAGAACAGTCAGAGAATGCCAAGCCAGTGGGAGGACTACTCAAG  
 AACCTGGAGAATCTGGTGGCTCAAACCAACTGGCCCTGTCTTGGCCAAAGGCTACCC

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HUMAN SAC CDNA SEQUENCE

**FIG. 3B**

ACCTGATGGCTTACGTCTGTATATTAAATGGGAGATGGGCAGAAATGTGGCCTTCTGAACACAGCCT  
TGCAGCTCTCTGAAACACAGGGAAATATACTGGAGAAATGCTGGCTGAACATGAACAAAGAATCATGGT  
ACTCAACCTCTGAGTTAAAGAAGACCAATGGCTTCAGACGATCTTGAGTCTCCATCATGGAAAAAA  
TTGTAGCAGGCAGGGTAAACATTCAAGGATCTTCAAAAAACAAATTCTGATGAGAGCTAATACCGTGG  
ACAATCATTCTAA

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Alignment of the rat and human sAC nucleotide sequences

**FIG. 4A**

hsAC	1	ATGAAACACTCCAAAAGAAGAATTCCAGGACTGGCCCATAGTCAGAATAGCAGCTCATTTACCAAGACCTCA
rsAC	1	ATGAGTGGCCCGAAGGCAGGAATTACAGGACAGGGCAATCGTCAAGATAGCTGCTCATTTACCGGACCTCA
hsAC	71	TTGTCTATGGACATTTCTCCCCAGAGCGACCCTTATGGAATTATTTGACGGAGTCCTGATGTTGTTGA
rsAC	71	TTGTCTATGGAGATTTCTCTCCCGAGCGGGCCGTCAAGTAAAATGTTGATGGAGTTCTGATGTTGTCGA
hsAC	141	TATTCAGGTTTACTGCAATGACTGAGAAGTTCAAGCACTGCCATGTACATGGACAGAGGGCTGAGCAG
rsAC	141	TATTCAGGTTTACTGCAATGACTGAGAAGTTCAAGCACTGCCATGTACATGGACAGAGGAGCCGAGCAG
hsAC	211	TTGGTGGAGATCCTCAACTACACATAAGTGCATAGTGGAGAAAGTGTGATTTGGAGGAGACATCC
rsAC	211	CTGGTGGAGATCCTCAACTACACATAAGTGCATAGTGGAGAAAGTACTGATTTGGAGGAGACATCC
hsAC	281	TGAAATTGCAGGTGATGCCACTGCTAGCCCTGTGGAGGGTGGACCGAAAGCAGCTGAAAAACATTATCAC
rsAC	281	TAATATTGCAGGTGACGCCCTGTTGCCCTGTGGAAAGTGGACGAAAGCAACTGAAGAATATCAC
hsAC	351	AGTGGTAATTAAATGTAGCCTGGAGATCCATGGATTGTTGAGACCCAGGAGTGGGAAGAAGGCCTAGAC
rsAC	351	GGTGGTAATTAAATGCAAGCCTGGAGATTCATGGCTTGTGAAGCCAAGGAGGTTGAAGAAGGCCTGGAT
hsAC	421	ATCCGAGTCAGATAGGACTGGCTGCTGCCACATCAGCATGTTGGCTTGGAGATGAAACACACAGCC
rsAC	421	ATTCGAGTTAAGATAGGACTGGCTGCTGCCACATCACCAGCATGTTGGCTTGGGATGAAACACAGGAAC
hsAC	491	ACTTCTGGTATTGGTCAGGCAGTGGACCGATGTGCGCCTTGCCAGAACATGGCTCAGATGAATGATGT
rsAC	491	ACTTCCTGGTATTGCCAAGGGTGGATGATGTACGCCCTGCTCAGAACATGGCTCAGATGAATGATGT
hsAC	561	TATTCTGTACCAAAACTGCTGGCAGCTCTGTGACCGGAGCATGATTGAAATTGAGAGTGTTCAGATCAG
rsAC	561	TATTCTGTACCAAAACTGCTGGCAGCTCTGTGATCGGAGCATGATTGAAATCGAGAGGATTCCGGATCAG
hsAC	631	AGAGCAGTTAAGGTTAACCTCTAAACCAACCCCCCAAATTAAATTGATGAATTTCACAAAGTGTAA
rsAC	631	AGAGCAGTTAAGGTTAGCTCTAAACCAACCCCCCAAACTTAAACTTCGACGAGTTTGTGCCAAGTGTAA
hsAC	701	CGACCTTCATGCATTATTATCCTCTGGTAGACACAAAAACCTCCTGAGGCTTGCAATGCACGGCTGAAGCC
rsAC	701	TGGCCTTCATGGATTATTATCCTCTGGTAGACACAAAAACCTCCTAAGGCTTGCAATGCATGCTGGAGTC
hsAC	771	TGATCCTGAACCTGGAGATGTCCTACAAAAGTATGTGATGGAAAGCATTTGAAGCAGATTGATAACAAA
rsAC	771	TGATCCTGAACCTGGAGTTGTCCTACAAAAGTATGTGATGGAAATCATTTGAAGCAGATTGATGACAAG
hsAC	841	CAGCTTCAGGGCTATTATCTGAGCTTCCGCCAGTGACCGATTGTGTTGTGAACCTGATGTTGAAGAC
rsAC	841	CAGCTTCAGGGCTATTATCTGAGCTTCCGCCAGTGACCGATTGTGTTGTGAACCTGATGTTAAAGAGC
hsAC	911	AAGACAAAGCAGAAGAGATAGGCCAGCCATCCAGGAATGCCTATATGCACATCACCTCTGTCCCTGAAGAAT
rsAC	911	AAGACAAAGCAGAAGTCATAGGATCAGCCATCCAAGCTGCCGTGTCACATCACCTCCGTCTGAAGGT
hsAC	981	CTTCCAGGCCAAATCAATAAGTCTTCATGTTGACAAGGGCTGCTCTTCCTCTGTCTTGGCTTC
rsAC	981	CTTCCAGGCCAGATCAATAAGGTCTTCATGTTGATAAGGGCTGCTCTTCCTCTGTCTTGGCTTC
hsAC	1051	CCTGGGAAAGGTACCTGACGAGCTCACTCATGCTCTGGAAATGTGCTATGGATATATTGACTTCTGCT
rsAC	1051	CCTGGGAAAGGCCCTGACGAGATCACTCACCGCTTGAAAGTGGCGTGGATATATTGACTTCTGCT
hsAC	1121	CTCAAGTCCACAAAATCCAAACTGTATCCATCGGTGTTGCCAGTGGGATTGTCTCTGTGGATCGTTGG
rsAC	1121	CTCAGGTCCACAAAATCCGTACTGTCTCCATCGGCGTCGCCAGTGGGATTGTCTCTGTGGATCGTTGG

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Alignment of the rat and human sAC nucleotide sequences

**FIG. 4B**

hsAC 1191	ACACACTGTGAGACACGAGTACACAGTCATTGGT	CAAAAAGTCAACTT	AGCTGCCAGGATGATGATGTAC
rsAC 1191	ACACACTGTGAGACACGAGTACACAGTCATTGGC	CAAAAGGTCAATATT	GCTGCCAGGATGATGATGTAT
hsAC 1261	TACCCAGGAATTGTGACCTGCCACTCTGTCAACCT	TACAATGGGAGCAACCT	ACCAGCGTACTTTTAAAG
rsAC 1261	TACCCAGGCATCGTGACCTGCCACTCTGTCAACATACGATGGC	AGCAACCTGCCAGCCT	ACTTTTAAAG
hsAC 1331	AGCTTCAAAGAAAGTTATGAAAGGT	GTTGCAGATTCTGGACCATTGTATCAGTATT	GGGGCGTACTGA
rsAC 1331	AGCTTCAAAGAAAGTCATGAAAGGA	GTTGCAGATCCCGGACCAGTGTATCAGTGTCTGGGCCTCAATGA	
hsAC 1401	GAAAGTCATTTGGTATGGCGTGCCTCATCTGCAACAGAAAGGAGGATT	ACCCCTTGCTGGGACGTAAT	
rsAC 1401	GAAAGTCATTTGGTATGGCCTATCTCATCTGCAACAGATATT	GAGGGCTACCCTTGCTGGGT	CGTGT
hsAC 1471	AAAGAGATCAAACATTTCATGT	TAATCTATGAAGAAATT	TTTGATATCTAACAGCAGCCAAAGTCTTAATGT
rsAC 1471	AGGGAGATCGACTATTTCATGT	TAATCTATGAAGGACTTTCTGATGACGAAC	TGCAGCCGAGTTCTAACATGT
hsAC 1541	ATGAGGGATTACCAGGATATGGAAAAAGCCAGA	ACTTATGAAAATT	GAGTACCTGGCC
rsAC 1541	ATGAAGGATTGCCAGGATATGGGAAAAGCCAGGT	ACTTATGAAATCGAGTAT	CTGGCCTCCCAGCATGA
hsAC 1608	GAATCACAGGATTATTGCCATTTCAT	TGAATAAGATCAGCTTCATCAAAC	TTTCTATACCATCCAGATG
rsAC 1611	GAACCATAGGGCTGTTGCTATTGCACTGACTAAGATCAGCTTCATCAA	AAATT	TTTACACTATCCAGATA
hsAC 1678	TTCATGGCCAATGTCTAGGCCTAGAC	ACTTGAAACATT	AAAGAACGACAGACCAAC
rsAC 1681	CTCATGGCTAACGTACTAGGTCTGGAT	ACTTGAAACATT	AAAGAACGACAGACCAAC
hsAC 1748	AAGTCATGACACTGTTGGATGAAAAGTTCTACTGTCT	CTTAATGACATT	TTCCATGTTAGTCCCTAT
rsAC 1751	GAGTCAAAGACGCTGTTGGATGATAAAATACCACTGTCT	CTTAACGACAT	TTCCATGTTAGTCCCGT
hsAC 1818	TTCTCGGGAGATTCCAGGATGAGCA	CCTTGAAAAGCAAAAACAATT	GGAAATTGTTATGAAGATC
rsAC 1821	TTCCCGGGAGATGTCCAGGATGAGCA	AGATAAGAAAGCAGAAGCAACT	GGAAAGCTCTGTTATGAAGATC
hsAC 1888	TTGAAAGCTGATAGTGAAGAGGAAAGGATTATT	TTTATCAT	TGAGGCCAGTTGTGGATTGACCT
rsAC 1891	CTGGAGCAAAACAGTGAGGGAGAAAGGATTAT	TTCATCATCGAC	GAGGCCAGTTGTGGACGTAGCCT
hsAC 1958	CCTGGAGATTATGGAGAAGCT	TATCCGGACTCTTCC	TATCTCATCATTATGTCCTGTGTCCTCGT
rsAC 1961	CCTGGGCCTTCATAGAAAAGCT	CATCCGGTCCATGCC	CATTGTTATGTCCTGTGTCCTCGC
hsAC 2028	TAACATTCCCTGTGCAGCTGCCAGGGCGTAAT	AAAGAACAGGAACACCACCTACAT	TGTCGTTGGTGCA
rsAC 2031	TGAAAACCTCCCTGCGCAGCTGCCAATGCCATAAT	GAAGAACGGAACACCACCTACAT	CACACTGGTAC
hsAC 2098	GTACAGCCTAACGACATCTCCAAACAAGATCTGT	CTTGACCTCAATGTGAGCT	GCATCTCCAAAGAACCTGG
rsAC 2101	ATGCAAGCCTCAGGAAATCGGGACAAGGTCTGT	GTTGACCTGAGTGTAAAGCAGCAT	CCCCAGAGAGCTTG
hsAC 2168	ACTCGTACCTGGGGAGGGAGCTGT	GGGATTCCATT	TGCTTAAAAACCTGGAACA
rsAC 2171	ACTCGTACCTGGTGGAGGGAGCTG	TTACTGTGAAGAAATT	GCTGAAAAACCTGACCCA
hsAC 2238	TCAATGAGGTACTCGTTTCCAACAAACGGAGT	CTGAGGAAAAGACAAAT	AGGACCTGGAATAACCTGTT
rsAC 2241	CCACAGAAATTCTCATTTCCAACAAAGCAGAGG	CTGAGGAAAAGACAAACGT	GACCTGGAATAACCTGTT

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Alignment of the rat and human sAC nucleotide sequences

**FIG. 4C**

hsAC 2308	AAGTATTCCATTAAAGCTAACAGAGAAGTAAACATGGTTACTCTCCATAGTGATAAGGAAAGTGAAGAAG
rsAC 2311	AAGTACTCTGTTAACGCCGACAGAAGACATGTATCTGTATACTTCC--ATAGCCGGGACAGAAAGAAG
hsAC 2378	TCTGTCACCTCACAAAGTGGCGCTCAGACTGAAAAACCTGTCACCTCCAACGTCATTAAAAGAAATCTCTCT
rsAC 2378	CCTGTTACCTTACAAGTGGTGTCAAGCTAAAGAACTTGTCAACCTCCAGCATCGCTCAAAGAAATCTCTCT
hsAC 2448	GATTCAGCTGGATAGCATGAGACTTTCCCAACCAAATGCTGGTGAGATGTGCTGCCATCATTGGCCTGACC
rsAC 2448	GGTTCAACTGGACAGCATGAGCCTTCCCATCAGATGCTGGTGACGTGTGCTGCTATCATTGGTCTAAC
hsAC 2518	TTCACCACTGAGTTGTTGTTGAGATTCTCCCTGTTGGAATATGAAGATGATGATCAAGACCCCTGGCAA
rsAC 2518	TTCACCAACAGAGCTGCTGTTGAGATTCTCCCTGCTGGAACATGAAGATGATGATCAAGGCCCTGGCCA
hsAC 2588	CCCTAGTGGAACTAACATTTTTATTGTTCCGGAATGGCAAGGAGCTTCAAAGGCCCTGAAACAGAA
rsAC 2588	CCCTAGTGGAACTAAATGTCTTGTGATTGCTTTCCGAGTAGCAAAGACCTTCAACTAGCCTTAAAGCAAAA
hsAC 2658	TGATCCCTCATTTGAGGTGCACATCGTTCCCTGTCTGAAGCCAGTGAAGGGATGGATCACGGTGAA
rsAC 2658	CGTGACCACTGTTGAAGTTCAATTATCGCTCTTGTCCCTGAAGTCCAAGGAAGGGTTAGCTTACAGTGAG
hsAC 2728	GAGGAACAGCTTCGTGAACCTGGAGAATGAGGTGATCGAGTGCACAGGATTGATTCTGTAACCCATATGA
rsAC 2728	GAGGAGCAGCTCCGTGAAATGGAAGGAGAGGTGATTGAATGCCCATCCTTCGGTTCTGCAGACCCATAA
hsAC 2798	TGCAGAAAACAGCCTACGAGCTGTCGGCTCAAGGACCAGAGAAAAGCCATGCACTTGAAATGTGCCCGCTT
rsAC 2798	TGCAGAAGACAGCCTACGAACTGTCGGCTCAAGGACCAGAGAAAAGCTTGCACTTGAAATGCCCGCTT
hsAC 2868	TTTAGAAGAAGATGCCACAGATGTGACCACTGCCGAGGCAGGACTTCATTCCCTATCATCACTTCACA
rsAC 2868	TTTGGAGGAGAGTGCCCATCGGTGCAACCACTGCAGAAACAGAGACTTCATTCCCTACCAACCACTTCATA
hsAC 2938	GTGAATTTCGGCTAACGCTTTAGACATGGATGCCATTAAAAGATGGCTATGTCTCATGGATTTAAA
rsAC 2938	GCGGACATTCAACTAACACTCTGGACATGGATACTGTCAAGAAGATGGTAAGTCCCACGGATTTAAA
hsAC 3008	CTGAAGAAAAGCTTATCTTGTCCAACCTCAGAGATTCCCTGAGACATCTGCATTTCCTGAAAATCGCAG
rsAC 3008	CTGAAGACGGAGGTCACTTTCTAAATCAGAGATCCCCAGGAAATTCAAATCTCCCCGAGAACATCAG
hsAC 3078	TCCTGAAGAAATAAGAGAAAAGATCTTGAATTCTTGTACCTTAAACAAAATGAAGACATCTGAC
rsAC 3075	CATCACAGAAACAAGGGAAAAATCTTGCATTCTTGTACAAATGTTATCATAAAGATGAGGACGTCTCAG
hsAC 3148	GAAGACATTATCCCTCTGGAATCTTGCAGTGTGAAAGAAATCCTAGAGATTGTCATCTGCCCTGGCCC
rsAC 3145	GATGATGTCACTCCCTCTAGAACATCGTGCCTTGTGAGGAGCTGCTCCAGATTGTCATCTGCCCTGGCCC
hsAC 3218	ACCATTTCTGGCTTTGGAGAAAATGACAAAGCCTTATATTACTCTTAGAAATTGCATCTGCTTATCT
rsAC 3215	AGCATTTCGTAGCCTTAAGAAGAAAACAACAAAGCCTTGTACTACTCCTAGAACATTGCATCTGCCATATCT
hsAC 3288	CATCTTTGTGATAACTACATGGCATACTGTATTGAAATGAAGGACAGAAGTTGCTAAAAACTCTCAAG
rsAC 3285	CATCCTGGAGACAACATAAACGCATACTGTATTGGCGAAGGGAAAGGCTGTTGAAATCTCTGACA
hsAC 3358	AAGGACAAAATCTTGGAGCCAGACATTGAGTCGCCACCTTTACAGCCTCAAAGGTGAGGTCTGTTCA
rsAC 3355	AATGAAGATTCTTGGAGTCAGACCTTGAATATGCTACGTTTATAGTCCTAAAGGTGAGATCTGTTTA

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Alignment of the rat and human sAC nucleotide sequences

**FIG. 4D**

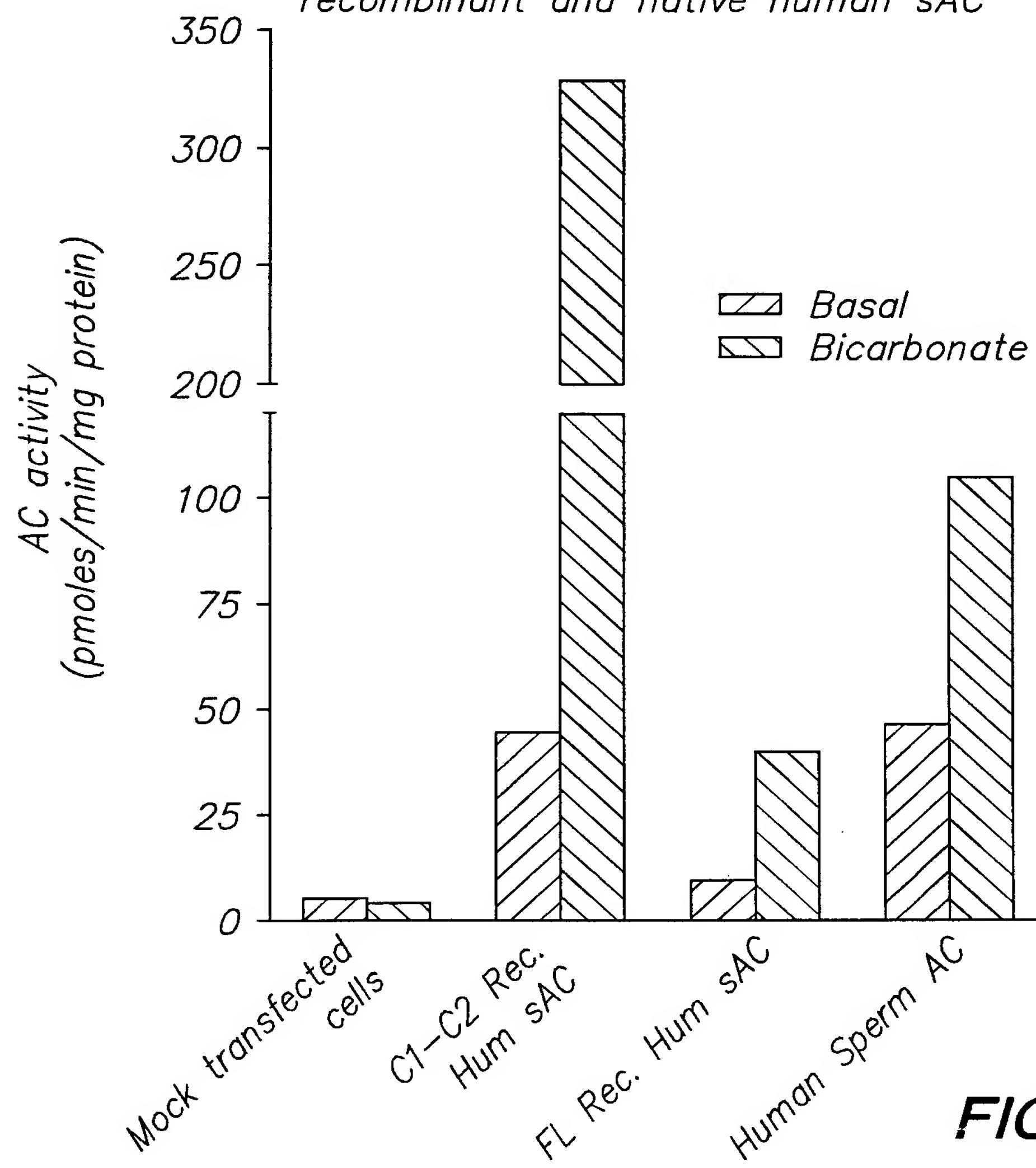
hsAC 3428	ATATGGGCCAGATAGTGCTTGCCAGAAAATGCTGAGGAAGGCACTGAAGCTCCTCAACCGAAATCTTCC
rsAC 3425	ATATGGGACAGATGGTGCTGCCAGAAAATGCTGAGAAAGGCACTGAAGCTCCTCAACAGAAATGTTCC
hsAC 3498	TITACAACCTTAATCTCCCTTGTTCTCCATATCCATGTCAGAAAAACAGACACTTTCATTTATGTAATCGG
rsAC 3495	CTGCAATCTACTCTCCCTGACTTTCCAAAATGCCACATTGAGAAAAACAGACTCTCCCACCTCATGAAACGAG
hsAC 3568	CAGGCCAAGAGAGGCCACCTCCAGGAAGAAGAGGCTGGCACAACTTTACCGGAAACACTGTCTGCCCTTT
rsAC 3565	CATIACCCAGGAGGGCTCGCTGCCAGGAAGAGCTGGCCAACTTTCCCTGCAAGTCGTCCTGCTTCT
hsAC 3638	CCTTGCTGTGGCGCATCTATAGCTACAGTTATCTTTTCACTGCAAGTATTATGCCAACCTGGCAGTTAT
rsAC 3632	CCCTGCTGTGGAAAGATCTATAGCTTGAACTTCTTTCCACTACAAGTACTATGTCGTCCTGGCAGCAAT
hsAC 3708	GATGCAAATGAATACTGCACTGGAAACTCAAAATTGTTCCAGATCATTAAGGCTTACCTAGACTATTCTG
rsAC 3702	AATGCAAGATGAACACCTCGTTAGAAACTCAAAACAATTTCCAGATCATCAAGGCTTTCCTGGACTTTTCC
hsAC 3778	CTATACCAACACCTGGCTGGTACAAAGGTGTGTGGTCAAATATGAAAGTCATGGCCATGGAGCACATCT
rsAC 3772	CTGTACCGCCATCTGGCTGGTACGAGGGCGTGTGGTCAAATATGAAATCCTGGTCAATGGAGCACT
hsAC 3848	TCAACCTCCCCCTGAAAGGCCAGGGCATTGAAATCGTGGCATACGTGGCTGAGACACTGGTCTTCACCAA
rsAC 3842	TGAAACCTCCCCCTGAAAGGCCAGGCATTGAAATCATGGCTATGCAGGCCACGGCACTGGGCCATATCAA
hsAC 3918	GCTCATTAATGGGACACCTGGATTGGCCATTGAGTTAGGCTCCCGAGCCCTCAAGATGTGGCACTGCTC
rsAC 3912	GTTCTTAACCGGTCACTGGACTTGGCCATTGAATTAGGCTCCCGAGCTCACAAAGATGTGGTCACTTCTC
hsAC 3988	CAGAATCCCAACCGACATTATCAGTCCTCTGCAGACTTAGCAGATGTCCTTCTGAACAGCAGATAAC
rsAC 3982	CGGAATCCCAACAAATACCATATGGTTCTCTGCAGACTGAGTAACACCTTTCTGAAGAGCAGATAAC
hsAC 4058	CGCAATTGATCCAGGTGCTGGGGCGGCTGTGGGAGCTTCTGTAACACAGGAACACATCTCAGCAAGGC
rsAC 4052	AGCATTGGTCCAGGTGCTGGATGGTTGTGGACCTTCTGTAACAGAGGACACATCTCAGCAAGGC
hsAC 4128	ATTTTCTATTTGTCTGCTTGGACATCCTGCTTATTCTGGTTTGTGTTATAGAACATTGAAGAATGT
rsAC 4122	ATTTTCTATTCGTCTGCTTGGACATCATGCTTATTCTGGCTTCATTACAGAACATTGAAGAATGT
hsAC 4198	TTGGAATTACACCAAATACGAAAACAACAGAACCTCAAGTCCACAGTGGACTCCTGGACTTT
rsAC 4192	TTGGAATTACACCCACAATGAAGACAACAGAACCTCAAGTCCAAAGCGGACTCCTGGACTTT
hsAC 4268	ATTCCCTCTGTAGCTATCTGGTATGCCAGACTTCAGGAATGGGACAACCTTACAAATTTCACATAGAGC
rsAC 4262	ACTCCTGCATAGCTGCTGGTACGCCAGACTTCAGGAATGGGACAACCTTACAAATTTCACATAGAGC
hsAC 4338	TAAAAAATCTTTGCCAAAGAACCATGACACTTACTTACTATGACGGAAATCTAGGTACATGGAGGGG
rsAC 4332	GAAGACTTTAGTGACTCGAAGAACCCCAACGGTCTTACTACGAAGGAATTCTAGGTATATGGAAGGG
hsAC 4408	CAAGTTCTTCACTTCAAAAAACAAATCAAAGAACAGTCAGAGAACGCCAGTGGGAGGAGCTAC
rsAC 4402	CAAGTCCTCICATCTTCAGAACAAATAGAACAGCAGGCCGAGAACATGCTCAGGACAGTGGGAGCTAC
hsAC 4478	TCAAGAACCTGGAGAATCTGGTGGCTAAAATACCACTGGCCCTGTCTTTGCCAAGGCTTACCCACCT
rsAC 4472	TTAAGGCCTTAGAGAACCCCTTGTGGCTAAAATACCACTGGCCCCGTCTTACCCAGGCTTACCCATT

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Alignment of the rat and human sAC nucleotide sequences

**FIG. 4E**

hsAC 4548	GATGGCCTAACGTCTGTATAATTAAATGGGAGATGGGCAGAAAATGTGGCCTCTTCCTGAACACAGCCTTGGG
rsAC 4542	GATGGCCTATGTCTGTATACTGATGGGAGACGGGCACAGTTGTGACTTCTTCCTAAACACAGCCTTGGAG
hsAC 4618	CTCTCTGAAACACAGGGGAATATACTGGAGAAATGCTGGCTGAAACATGAACAAAGAATCATGGTACTCAA
rsAC 4612	CTCTCTGAGACACAGGGGAATTGCTGGAGAAATGTTGGCTGAGCATGAGTAAGGAATGGTGGTACTCAG
hsAC 4688	CCTCTGAGTTAAAAAGAAGACCAATGGCTTCAGACGATCTTGAGTCTCCCATGATGGAAAAAATTGTAGG
rsAC 4682	CCCGCGAGTTGACAGGGAGATCAATGGCTTCAGACAGTCTTGAGTCTCCCATGGGGATAAAATTGTATC
hsAC 4758	AGGCAGGGTAAACATTCAAGGATGTTCAAAAAAACAAATTCTGATGAGAGCTAATAACCGTGACAATCAT
rsAC 4752	AGGCAACGTAACCCCTTCAGGATGTTCAAAAGAACAAATTCTGATGAGAGTTAAATATTCTGACAATCCT
hsAC 4828	TTCTAA-----
rsAC 4822	TTCTAATAATTATGAATGAGAACAAAGATTGCA

*Effect of Bicarbonate (50mM) on the recombinant and native human sAC*

**FIG. 5**

## SEQUENCE LISTING

&lt;110&gt; Conti, Marco

Jaiswal, Bijay Shankar

<120> Polynucleotides Encoding Human Soluble  
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of Use Thereof

&lt;130&gt; STAN-135WO

&lt;150&gt; 60/155,302

&lt;151&gt; 1999-09-21

&lt;150&gt; 60/191,327

&lt;151&gt; 2000-03-22

&lt;160&gt; 4

&lt;170&gt; FastSEQ for Windows Version 4.0

&lt;210&gt; 1

&lt;211&gt; 4833

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;400&gt; 1

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# INTERNATIONAL SEARCH REPORT

International application No.

PCT/US00/26129

## A. CLASSIFICATION OF SUBJECT MATTER

IPC(7) :C12Q 1/00; C12N 9/88, 1/20, 15/00; C07H 21/04; C07K 1/00  
US CL :424/94.5; 435/4, 232, 252.3, 320.1; 536/23.2; 530/350

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 424/94.5; 435/4, 232, 252.3, 320.1; 536/23.2; 530/350

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

West and STN - stn files included caplus, medline, biosis, embase, biotechds and scisearch

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A,P	US 6,001,553 A (BROACH ET AL.) 14 December 1999, see the entire document.	1-16
A,P	WICKER, R. et al. Cloning and expression of human adenylyl cyclase type VI in normal thyroid tissue. Biochimica et Biophysica Acta. 07 September 2000, Vol. 1493, pages 279-283, see the entire document.	1-16
Y	HELLEVUO, K. et al. The Characterization of a Novel Human Adenylyl Cyclase Which is Present in Brain and Other Tissues. The Journal of Biological Chemistry. 12 May 1995, Vol. 270, No. 19, pages 11581-11589, see the entire document.	1-16

Further documents are listed in the continuation of Box C.

See patent family annex.

• Special categories of cited documents:	"T"	later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
"A" document defining the general state of the art which is not considered to be of particular relevance	"X"	document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
"E" earlier document published on or after the international filing date	"Y"	document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	"&"	document member of the same patent family
"O" document referring to an oral disclosure, use, exhibition or other means		
"P" document published prior to the international filing date but later than the priority date claimed		

Date of the actual completion of the international search

14 NOVEMBER 2000

Date of mailing of the international search report

29 DEC 2000

Name and mailing address of the ISA/US  
Commissioner of Patents and Trademarks  
Box PCT  
Washington, D.C. 20231

Facsimile No. (703) 305-3230

Authorized officer

*Dorethea Lawrence* *for*  
TEKCHAND SAIDHA

Telephone No. (703) 308-0196

# INTERNATIONAL SEARCH REPORT

International application No.

PCT/US00/26129

## Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This international report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1.  **Claims Nos.:**  
because they relate to subject matter not required to be searched by this Authority, namely:
  
2.  **Claims Nos.:**  
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
  
3.  **Claims Nos.:**  
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

## Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

Please See Extra Sheet.

1.  As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2.  As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3.  As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
  
4.  No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

1-16

Remark on Protest

The additional search fees were accompanied by the applicant's protest.

No protest accompanied the payment of additional search fees.

# INTERNATIONAL SEARCH REPORT

International application No.

PCT/US00/26129

## BOX II. OBSERVATIONS WHERE UNITY OF INVENTION WAS LACKING

This ISA found multiple inventions as follows:

This application contains the following inventions or groups of inventions which are not so linked as to form a single inventive concept under PCT Rule 13.1. In order for all inventions to be searched, the appropriate additional search fees must be paid.

Group I, claim(s)1-16, drawn to a polypeptide (soluble human adenylyl cyclase activity), composition comprising the polypeptide and polynucleotide encoding the polypeptide, vector and host cell.

Group II, claim(s) 17-19, drawn to antibody.

Group III, claim(s) 20-22, drawn to a method of identifying a substance which modulates soluble human adenylyl cyclase.

Group IV, claims 23-29, drawn to a method of inhibiting spermatogenesis in a male, administering a substance that inhibit soluble adenylyl cyclase activity.

The inventions listed as Groups I-IV do not relate to a single inventive concept under PCT Rule 13.1 because, under PCT Rule 13.2, they lack the same or corresponding special technical features for the following reasons: Group I has a special technical feature of nucleotide sequence encoding human adenylyl cyclase which Groups II-IV do not share; Group II has a special technical feature of an antibody which Groups I & III-IV do not share; Group III has a special technical feature of a method of identification of a substance that modulate human adenylyl cyclase which Groups I-II & IV do not share; Group IV has a special technical feature of a method of inhibiting spermatogenesis in a male by inhibiting human adenylyl cyclase which Groups I-III do not share.